

Performance of the UV24 Unit Against Zoonotic Pathogens

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For Medical Illumination International

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Introduction

The UV24 unit disinfects airborne pathogens using ultraviolet light and employs an airflow of 50 cfm. It is intended for applications in rooms occupied by humans and can also be applied to veterinary environments for the control of zoonotic pathogens. The unit can protect human occupants from zoonotic diseases in rooms that house various types of animals and can protect animals from zoonotic diseases that hail from other animals. The animals for which it is intended to offer protection from airborne diseases include dogs, cats, poultry & birds, rodents and hamsters, horses, swine, cows, rabbits, lizards and snakes. The pathogens that have been identified as being of greatest concern in the veterinary industry include but are not limited to *Microsporium canis*, Canine Distemper virus, Canine Influenza (H2N7), Feline Infectious peritonitis, Feline coronavirus, *Cryptococcus* spp., Feline Rhinotracheitis, Reovirus, *Micrococcus*, *Staphylococcus aureus* (MRSA), *Pasteurella multocida*, Porcine circovirus, and *Bordetella pertussis*. A complete list of potentially airborne zoonotic pathogens is included for which the UV susceptibility is known and the predicted performance of the UV24 unit against these pathogens is evaluated. Also evaluated are the removal rates of pathogens in a model room of 800 ft³ volume with no outside air. Included for completeness in Appendix A is a list of all zoonotic pathogens which are not airborne.

Evaluation of Select Zoonotic Pathogens

Table 1 summarizes critical information for the aforesaid zoonotic pathogens including ultraviolet (UV) susceptibility, the primary diseases caused, and transmissivity to humans. The affected animal species is identified with a single letter code identified below the table. The Group identifies Viruses (V), Bacteria (B), Fungi (F) and Fungal Spores (FS) with a single letter code. Only one of these select pathogens, *Pasteurella multocida*, has no known UV susceptibility. The UV susceptibilities of the remaining pathogens are taken from Kowalski (2009).

The UV24 unit has an airflow of 50 cfm and produces a UV Dose of 198 J/m². It also employs a MERV 6 filter. The performance curve for the MERV 6 filter model is based on Kowalski et al (1999) and has been extended into the virus size range (below about 0.1 micron) as shown in Figure 1. The mean diameters of the subject microbes have been used to predict the filter removal rates.

Table 1: Select Airborne Zoonotic Pathogens

| PATHOGEN | HOST | GROUP | DISEASE | Mean dia. μm | UV k m^2/J | UV D90 J/m^2 | Trans to Human |
|--------------------------------------------|-------------|--------------|----------------------------------|-------------------------------------------|----------------------------------------------|------------------------------------------------|-----------------------|
| Bordetella bronchiseptica | DCR | B | Kennel cough | 0.707 | 0.0364 | 63.258 | Yes |
| Canine Distemper Virus (CDV) | DC | V | Canine distemper | 0.15 | 0.11 | 20.933 | No |
| Canine Influenza H3N2 | DC | V | Flu | 0.098 | 0.101 | 22.798 | No |
| Canine Influenza H3N8 | DE | V | Flu | 0.098 | 0.101 | 22.798 | No |
| Coronavirus | P | V | Infectious bronchitis | 0.113 | 0.01 | 230.26 | Yes |
| Cryptococcus farciminosus | E | F | Cryptococcosis | 4.9 | 0.0167 | 137.88 | Yes |
| Cryptococcus neoformans | DCBEO | F | Cryptococcosis | 4.899 | 0.0167 | 137.88 | Yes |
| Feline Herpesvirus T1 (FVR) | C | V | Colds, fever, rhinotracheitis | 0.18 | 0.1046 | 22.013 | No |
| Feline Infectious Peritonitis virus (FIP) | C | V | Peritonitis, Enteric Coronavirus | 0.11 | 0.0053 | 434.45 | No |
| Micrococcus spp. | P | B | various | 1.2 | 0.0298 | 77.164 | Yes |
| Microsporium canis | DCREO | FS | Dermatophyopsis, ringworm | 2.96 | 0.0096 | 240 | Yes |
| Pasteurella multocida | DCBPRSO | B | Pasteurellosis, fowl cholera | 0.6 | unk | unk | Yes |
| Psittacine Beak and Feather Disease (PBFD) | B | V | Beak & feather infections | 0.055 | 0.007 | 328.94 | No |
| Reovirus | BR | V | Colds, fever, pneumonia | 0.08 | 0.016 | 143.91 | Yes |
| Staphylococcus aureus (MRSA) | DCBPRES | B | MRSA, various infections | 0.866 | 0.596 | 3.8634 | Yes |

D = Dog
 C = Cat
 R = Rodent
 E = Equine
 O = Bovine
 P = Poultry, Birds
 S = Swine

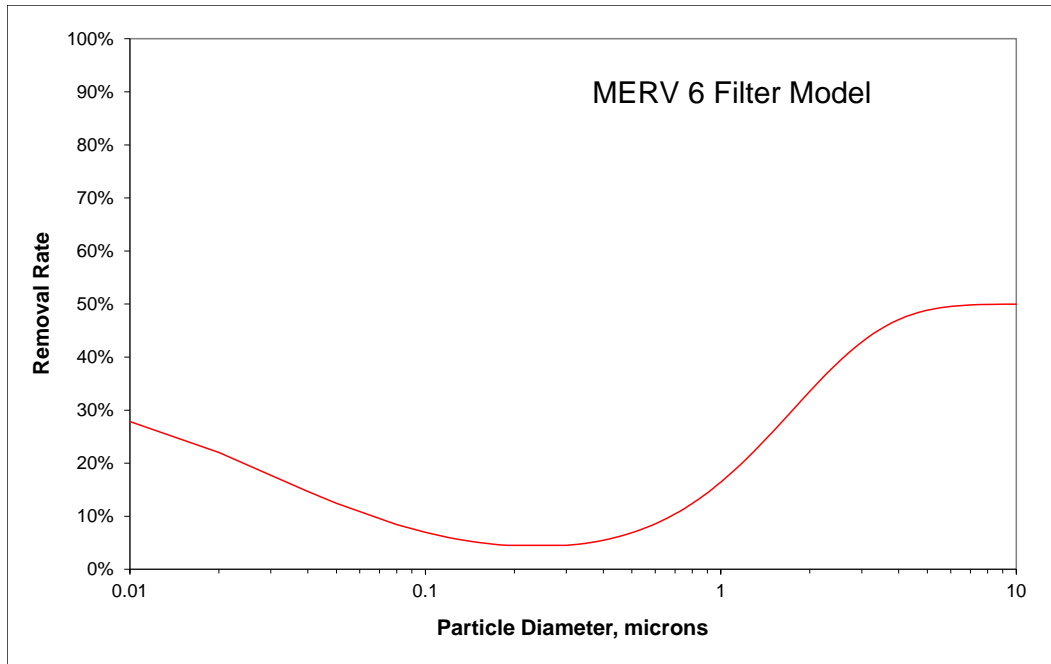


Figure 1: MERV 6 Filter Model extended into the virus size range. Per Kowalski et al (1999).

Table 2 summarizes the select pathogens from Table 1 and computes their removal rates from both the MERV 6 filter and the UV lamps, which produce a UV Dose of 198 J/m². The final column shows the total removal rate in a single pass through the UV24 unit.

Figure 2 illustrates the reduction over time of airborne pathogens in a room of 800 ft³ volume with no outside air. The initial concentration is assumed to be 1000 CFU/ft³. The removal rates represent the combined removal rates of UV and filtration as indicated in Table 2A.

The pathogens in Table 2 are repeated in the following sections where they are analyzed as a group for the subject host animals.

Table 2: UV24 Kill Rates for Select Airborne Zoonotic Pathogens

| PATHOGEN | Mean | MERV 6 | UV k | Single Pass | Total Removal |
|--------------------------------------------|--------------------|---------------|-----------------------|-----------------|---------------|
| | Dia. μm | Filtration, % | m^2/J | UV Kill Rate, % | Rate, % |
| Bordetella bronchiseptica | 0.707 | 10.57 | 0.0364 | 99.93 | 99.93 |
| Canine Distemper Virus (CDV) | 0.15 | 5.09 | 0.11 | 100.00 | 100.00 |
| Canine Influenza H3N2 | 0.098 | 7.09 | 0.101 | 100.00 | 100.00 |
| Canine Influenza H3N8 | 0.098 | 7.09 | 0.101 | 100.00 | 100.00 |
| Coronavirus | 0.113 | 6.29 | 0.01 | 86.19 | 87.06 |
| Cryptococcus farciminosus | 4.9 | 48.72 | 0.0167 | 96.34 | 98.12 |
| Cryptococcus neoformans | 4.9 | 48.72 | 0.0167 | 96.34 | 98.12 |
| Feline Herpesvirus T1 (FVR) | 0.18 | 4.60 | 0.1046 | 100.00 | 100.00 |
| Feline Infectious Peritonitis virus (FIP) | 0.11 | 6.43 | 0.0053 | 64.99 | 67.24 |
| Micrococcus spp. | 1.2 | 20.41 | 0.02984 | 99.73 | 99.78 |
| Microsporium canis | 2.96 | 42.60 | 0.009594 | 85.04 | 91.41 |
| Pasteurella multocida | 0.6 | 8.58 | unk | unk | 8.58 |
| Psittacine Beak and Feather Disease (Pbfd) | 0.055 | 11.55 | 0.007 | 74.99 | 77.88 |
| Reovirus | 0.08 | 8.44 | 0.016 | 95.79 | 96.15 |
| Staphylococcus aureus (MRSA) | 0.866 | 13.72 | 0.596 | 100.00 | 100.00 |

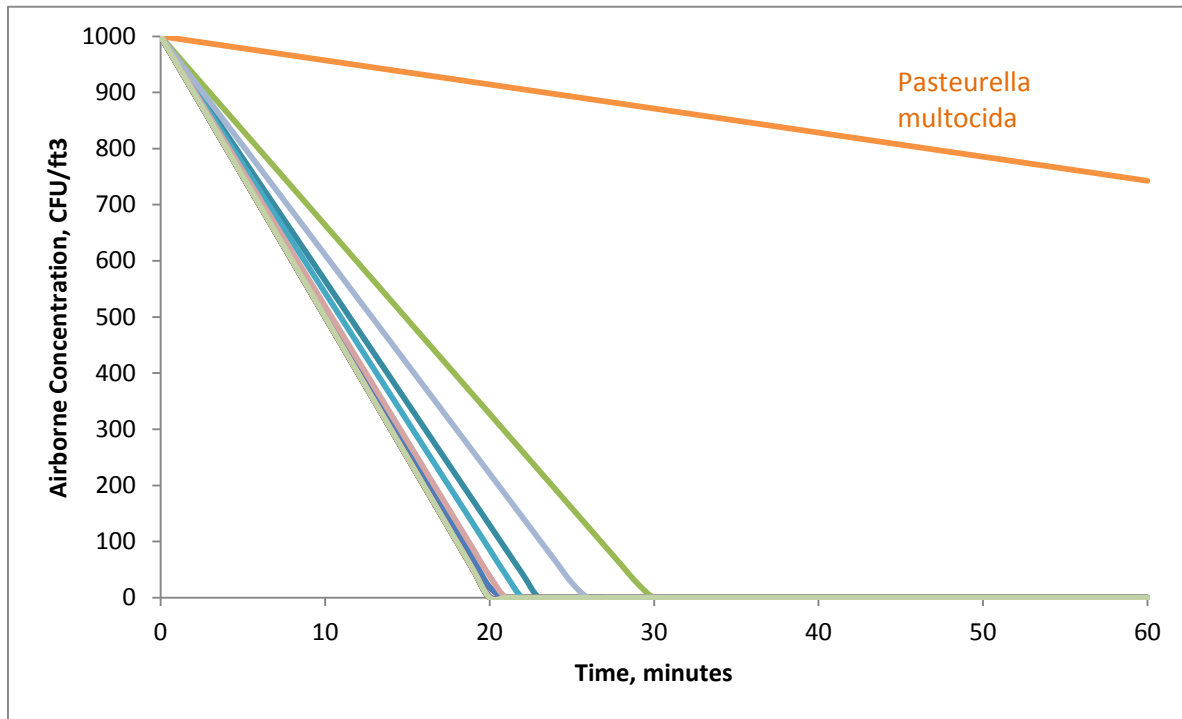


Figure 2: Removal of airborne pathogens from a room of 800 ft³ volume by the UV24 unit. *Pasteurella multocida* has no known UV susceptibility and this curve represents filtration only.

Airborne Diseases of Dogs

Canine diseases form the largest class of pathogens and include many that are transmissible to humans by contact and via the airborne route. Table 3 summarizes all the canine pathogens that are known or suspected to be transmissible by the airborne route. Some of these pathogens do not have a known UV rate constant but they are included in the tables for completeness and only the filter removal rate is given. Table 3A summarizes the calculated removal rates for all the pathogens in Table 3, except where the UV rate constant is unknown. Figure 3 illustrates the removal rates of the pathogens in Table 3A in a model room of 800 ft³ with no outside air and this graph only includes those pathogens with known UV rate constants. It can be observed in Figure 3 that all pathogens are removed to near-zero levels within about one hour.

Table 3: Potentially Airborne Dog Pathogens and Allergens

| PATHOGEN | GROUP | DISEASE | Mean | UV k | UV D90 | Trans to Human |
|-------------------------------------|-----------------|--------------------------------|--------------------|-----------------------|-----------------------|----------------|
| | | | Dia. μm | m^2/J | J/m^2 | |
| Aspergillus spp. | Fungal Spore | Aspergillosis | 3.354 | 0.00058 | 3970 | Yes |
| Avian Influenza A | Virus | flu | 0.098 | 0.101 | 23 | Yes |
| Bacillus anthracis | Bacterial Spore | Anthrax | 1.118 | 0.02702 | 85 | Yes |
| Blastomyces dermatitidis | Fungal Spore | dermatitis | 11 | 0.0165 | 140 | Yes |
| Bordetella bronchiseptica | Bacteria | kennel cough | 0.707 | 0.0364 | 63 | Yes |
| Brachyspira pilosicoli | Bacteria | Gastrointestinal illness | 0.5 | 0.0324 | 71 | No |
| Brucella abortus | Bacteria | Brucellosis, undulant fever | 0.57 | 0.0307 | 75 | Yes |
| Brucella canis | Bacteria | Brucellosis, undulant fever | 0.566 | 0.048 | 48 | Yes |
| Brucella melitensis | Bacteria | Brucellosis | 0.566 | 0.047 | 49 | Yes |
| Brucella suis | Bacteria | Brucellosis | 0.57 | 0.047 | 49 | Yes |
| Burkholderia mallei | Bacteria | Glanders | 0.77 | 0.034 | 68 | Yes |
| Burkholderia pseudomallei | Bacteria | Melioidosis | 0.494 | 0.0344 | 67 | Yes |
| Campylobacter coli | Bacteria | Enteritis, Campylobacteriosis | 2.12 | 0.144 | 16 | Yes |
| Campylobacter jejuni | Bacteria | Enteritis | 2.12 | 0.1444 | 16 | Yes |
| Canine Adenovirus Type 1 (CAv-1) | Virus | hepatitis | 0.093 | 0.026 | 89 | No |
| Canine Adenovirus Type 2 (CAv-2) | Virus | kennel cough | 0.093 | 0.018 | 128 | No |
| Canine Calicivirus (CaCV) | Virus | fever, conjunctivitis | 0.034 | 0.0345 | 67 | No |
| Canine Coronavirus | Virus | colds | 0.113 | 0.377 | 6 | No |
| Canine Distemper Virus (CDV) | Virus | Canine distemper | 0.15 | 0.11 | 21 | No |
| Canine Influenza H3N2 | Virus | flu | 0.098 | 0.101 | 23 | No |
| Canine Influenza H3N8 | Virus | flu | 0.098 | 0.101 | 23 | No |
| Canine Norovirus | Virus | gastroenteritis | 0.035 | 0.0304 | 76 | No |
| Canine Parvovirus 2 | Virus | Intestinal, cardiac disease | 0.022 | 0.092 | 25 | No |
| Chlamydomphila pneumoniae | Bacteria | Chlamydia | 0.283 | 0.039 | 59 | Yes |
| Clostridium botulinum | Bacteria | Botulism | 1.975 | 0.0412 | 56 | Yes |
| Clostridium perfringens | Bacteria | sepsis, toxins, food poisoning | 5 | 0.06 | 38 | Yes |
| Clostridium tetani | Bacteria | tetanus | 5 | 0.04699 | 49 | Yes |
| Coccidioides immitis | Fungal Spore | Coccidioidomycosis | 3.464 | - | - | Yes |
| Coxiella burnetii | Bacteria | Q Fever | 0.283 | 0.1535 | 15 | Yes |
| Cryptococcus neoformans | Fungi | Cryptococcosis | 4.899 | 0.0167 | 138 | Yes |
| Dermatophilus congolensis | Bacteria | dermatophilosis, mud fever | 1 | - | - | No |
| Epidermophyton | Fungi | Dermatophytosis | 8.5 | - | - | Yes |
| Erysipelothrix spp. | Bacteria | Erysipeloid | 1 | - | - | Yes |
| Feline Influenza A (H7N2) | Virus | Flu | 0.10 | 0.101 | 23 | Yes |
| Francisella tularensis | Bacteria | tularemia, pneumonia, fever | 0.2 | 0.009 | 256 | Yes |
| Hantavirus (Hantaan Virus) | Virus | Hemorrhagic Fever | 0.095 | 0.0688 | 33 | Yes |
| Hendra Virus | Virus | Pneumonia | 0.175 | 0.230259 | 10 | Yes |
| Histoplasma capsulatum | Fungal Spore | URD | 2.236 | 0.01645 | 140 | Yes |
| Influenza A virus | Virus | flu, secondary pneumonia | 0.098 | 0.101 | 23 | Yes |
| Leptospira spp. | Spirochete | Leptospirosis | 0.1 | NA | - | Yes |
| Listeria monocytogenes | Bacteria | Listeriosis | 0.707 | 0.0127 | 181 | Yes |
| Louping III (LIV) | Virus | Encephalomyelitis | 0.05 | 0.003289 | 700 | Yes |
| Lymphocytic choriomeningitis (LCMV) | Virus | Armstrong's disease | 0.087 | 0.0605 | 38 | Yes |
| Microsporium spp. | Fungal Spore | Dermatophytosis | 2.96 | - | - | Yes |
| Mycobacterium bovis | Bacteria | Tuberculosis | 0.637 | 0.181 | 13 | Yes |
| Mycobacterium tuberculosis | Bacteria | Tuberculosis | 0.637 | 0.472 | 5 | Yes |
| Nipah virus (Henipah virus) | Virus | respiratory syndrome | 0.175 | 0.328941 | 7 | No |
| Parainfluenza virus | Virus | flu, colds, croup, pneumonia | 0.194 | 0.1086 | 21 | Yes |
| Pneumocystis carinii | Fungal Spore | pneumocystosis | 2 | - | - | Yes |
| Pseudorabies (PRV) | Virus | Aujeszky's Disease Virus (ADV) | 0.194 | 0.0676 | 34 | No |
| Rabies virus | Virus | rabies | 0.07 | 0.219 | 11 | Yes |
| Spirillum minus | Bacteria | Rat Bite Fever | 1 | - | - | No |
| Sporothrix schenckii | Fungal Spore | Sporotrichosis | 6.325 | - | - | Yes |
| Staphylococcus aureus (MRSA) | Bacteria | MRSA, various infections | 0.866 | 0.596 | 4 | Yes |
| Streptobacillus moniliformis | Bacteria | Rat Bite Fever | 0.707 | - | - | No |
| Streptococcus pyogenes | Bacteria | fever | 0.894 | 1.561 | 1 | Yes |
| Swine Influenza | Virus | H1N1 flu | 0.1 | 0.098 | 23 | Yes |
| Trichophyton spp. | Fungal Spore | Dermatophytosis | 7 | 0.00411 | 560 | Yes |
| Yersinia pestis | Bacteria | Bubonic & Pneumonic Plague | 0.707 | 0.106 | 22 | Yes |

Table 3A: UV24 Removal Rates for Dog Pathogens

| PATHOGEN | Mean | MERV 6 | UV k | Single Pass | Total Removal |
|-------------------------------------|--------------------|---------------|-----------------------|-----------------|---------------|
| | Dia. μm | Filtration, % | m^2/J | UV Kill Rate, % | Rate, % |
| Aspergillus spp. | 3.354 | 44.76 | 0.00058 | 10.85 | 50.75 |
| Avian Influenza A | 0.098 | 7.09 | 0.101 | 100.00 | 100.00 |
| Bacillus anthracis | 1.118 | 18.79 | 0.02702 | 99.53 | 99.61 |
| Blastomyces dermatitidis | 11 | 50.00 | 0.0165 | 96.19 | 98.09 |
| Bordetella bronchiseptica | 0.707 | 10.57 | 0.0364 | 99.93 | 99.93 |
| Brachyspira pilosicoli | 0.5 | 6.89 | 0.0324 | 99.84 | 99.85 |
| Brucella abortus | 0.57 | 8.05 | 0.0307 | 99.77 | 99.79 |
| Brucella canis | 0.566 | 7.98 | 0.048 | 99.99 | 99.99 |
| Brucella melitensis | 0.566 | 7.98 | 0.047 | 99.99 | 99.99 |
| Brucella suis | 0.57 | 8.05 | 0.047 | 99.99 | 99.99 |
| Burkholderia mallei | 0.77 | 11.80 | 0.034 | 99.88 | 99.89 |
| Burkholderia pseudomallei | 0.494 | 6.80 | 0.0344 | 99.89 | 99.90 |
| Campylobacter coli | 2.12 | 35.03 | 0.144 | 100.00 | 100.00 |
| Campylobacter jejuni | 2.12 | 35.03 | 0.1444 | 100.00 | 100.00 |
| Canine Adenovirus Type 1 (CAV-1) | 0.093 | 7.41 | 0.026 | 99.42 | 99.46 |
| Canine Adenovirus Type 2 (CAV-2) | 0.093 | 7.41 | 0.018 | 97.17 | 97.38 |
| Canine Calicivirus (CaCV) | 0.034 | 16.39 | 0.0345 | 99.89 | 99.91 |
| Canine Coronavirus | 0.113 | 6.29 | 0.377 | 100.00 | 100.00 |
| Canine Distemper Virus (CDV) | 0.15 | 5.09 | 0.11 | 100.00 | 100.00 |
| Canine Influenza H3N2 | 0.098 | 7.09 | 0.101 | 100.00 | 100.00 |
| Canine Influenza H3N8 | 0.098 | 7.09 | 0.101 | 100.00 | 100.00 |
| Canine Norovirus | 0.035 | 16.08 | 0.0304 | 99.76 | 99.80 |
| Canine Parvovirus 2 | 0.022 | 21.04 | 0.092 | 100.00 | 100.00 |
| Chlamydomydia pneumoniae | 0.283 | 4.42 | 0.039 | 99.96 | 99.96 |
| Clostridium botulinum | 1.975 | 33.20 | 0.0412 | 99.97 | 99.98 |
| Clostridium perfringens | 5 | 48.83 | 0.06 | 100.00 | 100.00 |
| Clostridium tetani | 5 | 48.83 | 0.04699 | 99.99 | 100.00 |
| Coccidioides immitis | 3.464 | 45.25 | - | - | - |
| Coxiella burnettii | 0.283 | 4.42 | 0.1535 | 100.00 | 100.00 |
| Cryptococcus neoformans | 4.899 | 48.72 | 0.0167 | 96.34 | 98.12 |
| Dermatophilus congolensis | 1 | 16.43 | - | - | - |
| Epidermophyton | 8.5 | 49.96 | - | - | - |
| Erysipelothrix spp. | 1 | 16.43 | - | - | - |
| Feline Influenza A (H7N2) | 0.098 | 7.09 | 0.101 | 100.00 | 100.00 |
| Francisella tularensis | 0.2 | 4.43 | 0.009 | 83.17 | 83.91 |
| Hantavirus (Hantaan Virus) | 0.095 | 7.28 | 0.0688 | 100.00 | 100.00 |
| Hendra Virus | 0.175 | 4.67 | 0.2303 | 100.00 | 100.00 |
| Histoplasma capsulatum | 2.236 | 36.37 | 0.0165 | 96.15 | 97.55 |
| Influenza A virus | 0.098 | 7.09 | 0.1010 | 100.00 | 100.00 |
| Leptospira spp. | 0.1 | 6.97 | - | - | - |
| Listeria monocytogenes | 0.707 | 10.57 | 0.0127 | 91.91 | 92.77 |
| Louping Ill (LIV) | 0.05 | 12.45 | 0.0033 | 47.86 | 54.35 |
| Lymphocytic choriomeningitis (LCMV) | 0.087 | 7.85 | 0.0605 | 100.00 | 100.00 |
| Microsporium spp. | 2.96 | 42.60 | - | - | - |
| Mycobacterium bovis | 0.637 | 9.25 | 0.181 | 100.00 | 100.00 |
| Mycobacterium tuberculosis | 0.637 | 9.25 | 0.472 | 100.00 | 100.00 |
| Nipah virus (Henipah virus) | 0.175 | 4.67 | 0.328940728 | 100.00 | 100.00 |
| Parainfluenza virus | 0.194 | 4.47 | 0.1086 | 100.00 | 100.00 |
| Pneumocystis carinii | 2 | 33.53 | - | - | - |
| Pseudorabies (PRV) | 0.194 | 4.47 | 0.0676 | 100.00 | 100.00 |
| Rabies virus | 0.07 | 9.46 | 0.219 | 100.00 | 100.00 |
| Spirillum minus | 1 | 16.43 | - | - | - |
| Sporothrix schenckii | 6.325 | 49.66 | - | - | - |
| Staphylococcus aureus (MRSA) | 0.866 | 13.72 | 0.596 | 100.00 | 100.00 |
| Streptobacillus moniliformis | 0.707 | 10.57 | - | - | - |
| Streptococcus pyogenes | 0.894 | 14.28 | 1.561 | 100.00 | 100.00 |
| Swine Influenza | 0.1 | 6.97 | 0.098 | 100.00 | 100.00 |
| Trichophyton spp. | 7 | 49.82 | 0.00411 | 55.68 | 77.76 |
| Yersinia pestis | 0.707 | 10.57 | 0.106 | 100.00 | 100.00 |

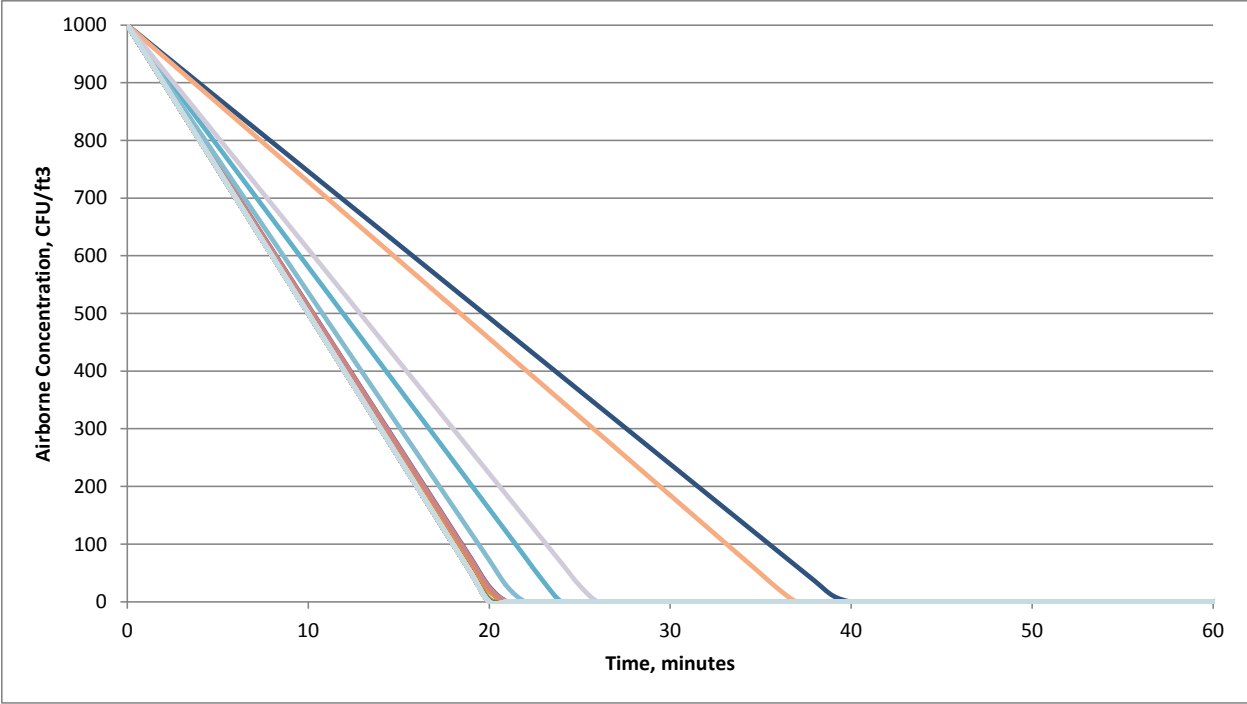


Figure 3: Removal of Canine pathogens by the UV24 unit in a room of 800 ft³ volume with no outside air. Only microbes from Table 3A with known UV rate constants are included.

Airborne Diseases of Cats

Feline diseases form the next largest group of zoonotic diseases and include many that are potentially airborne and that can transmit to humans. Table 4 summarizes all of the cat pathogens and allergens that are known or suspected to transmit by the airborne route.

Table 4: Potentially Airborne Cat Pathogens and Allergens

| PATHOGEN | GROUP | DISEASE | Mean | UV k | UV D90 | Trans to Human |
|-------------------------------------------|-----------------|-----------------------------------|--------------------|-----------------------|-----------------------|----------------|
| | | | Dia. μm | m^2/J | J/m^2 | |
| Aspergillus spp. | Fungal Spore | Aspergillosis | 3.354 | 0.00058 | 3970 | Yes |
| Avian Influenza A | Virus | flu | 0.098 | 0.101 | 23 | Yes |
| Bacillus anthracis | Bacterial Spore | Anthrax | 1.118 | 0.02702 | 85 | Yes |
| Bordetella bronchiseptica | Bacteria | kennel cough | 0.707 | 0.0364 | 63 | Yes |
| Brachyspira pilosicoli | Bacteria | Gastrointestinal illness | 0.5 | 0.0324 | 71 | No |
| Burkholderia mallei | Bacteria | Glanders | 0.77 | 0.034 | 68 | Yes |
| Burkholderia pseudomallei | Bacteria | Melioidosis | 0.494 | 0.0344 | 67 | Yes |
| Campylobacter coli | Bacteria | Enteritis, Campylobacteriosis | 2.12 | 0.144 | 16 | Yes |
| Campylobacter jejuni | Bacteria | Enteritis | 2.12 | 0.1444 | 16 | Yes |
| Canine Distemper Virus (CDV) | Virus | Canine distemper | 0.15 | 0.11 | 21 | No |
| Canine Influenza H3N2 | Virus | flu | 0.098 | 0.101 | 23 | No |
| Canine Parvovirus 2 | Virus | Intestinal, cardiac disease | 0.022 | 0.092 | 25 | No |
| Chlamydomyces felis | Bacteria | Chlamydia | 0.283 | 0.0384 | 60 | No |
| Clostridium botulinum | Bacteria | Botulism | 1.975 | 0.0412 | 56 | Yes |
| Clostridium tetani | Bacteria | tetanus | 5 | 0.04699 | 49 | Yes |
| Coccidioides immitis | Fungal Spore | Coccidioidomycosis | 3.464 | - | - | Yes |
| Cowpox | Virus | cow pox | 0.173 | 0.135 | 17 | Yes |
| Coxiella burnetii | Bacteria | Q Fever | 0.283 | 0.1535 | 15 | Yes |
| Cryptococcus neoformans | Fungi | Cryptococcosis | 4.899 | 0.0167 | 138 | Yes |
| Epidermophyton | Fungi | Dermatophytosis | 8.5 | - | - | Yes |
| Erysipelothrix spp. | Bacteria | Erysipeloid | 1 | - | - | Yes |
| Feline Calicivirus (FeCV) | Virus | URD, pneumonia | 0.037 | 0.0345 | 67 | No |
| Feline Distemper (Feline Parvovirus, FPV) | Virus | Distemper, Panleukopenia | 0.022 | 0.092 | 25 | No |
| Feline Herpesvirus T1 (Rhinitis, FVR) | Virus | colds, fever | 0.18 | 0.1046 | 22 | No |
| Feline Infectious Peritonitis virus (FIP) | Virus | Peritonitis, Enteric Coronavirus | 0.11 | 0.0053 | 434 | No |
| Feline Influenza A (H7N2) | Virus | Flu | 0.10 | 0.101 | 23 | Yes |
| Feline Panleukopenia (Picornavirus) (FPV) | Virus | Panleukopenia | 0.037 | 0.066 | 35 | No |
| Francisella tularensis | Bacteria | tularemia, pneumonia, fever | 0.2 | 0.009 | 256 | Yes |
| Hantavirus (Hantaan Virus) | Virus | Hemorrhagic Fever | 0.095 | 0.0688 | 33 | Yes |
| Hendra Virus | Virus | Pneumonia | 0.175 | 0.230259 | 10 | Yes |
| Histoplasma capsulatum | Fungal Spore | URD | 2.236 | 0.01645 | 140 | Yes |
| Influenza A virus | Virus | flu, secondary pneumonia | 0.098 | 0.101 | 23 | Yes |
| Leptospira spp. | Spirochete | Leptospirosis | 0.1 | NA | - | Yes |
| Listeria monocytogenes | Bacteria | Listeriosis | 0.707 | 0.0127 | 181 | Yes |
| Microsporium spp. | Fungal Spore | Dermatophytosis | 2.96 | - | - | Yes |
| Mycobacterium avium | Bacteria | Paratuberculosis, Johne's Disease | 1.118 | 0.04387 | 52 | Yes |
| Mycobacterium bovis | Bacteria | Tuberculosis | 0.637 | 0.181 | 13 | Yes |
| Mycobacterium tuberculosis | Bacteria | Tuberculosis | 0.637 | 0.472 | 5 | Yes |
| Mycoplasma spp. | Bacteria | infectious anemia, eye disease | 0.177 | 0.284 | 8 | Yes |
| Nipah virus (Henipah virus) | Virus | respiratory syndrome | 0.175 | 0.328941 | 7 | No |
| Pneumocystis carinii | Fungal Spore | pneumocystosis | 2 | - | - | Yes |
| Pseudorabies (PRV) | Virus | Aujeszky's Disease Virus (ADV) | 0.194 | 0.0676 | 34 | No |
| Rabies virus | Virus | rabies | 0.07 | 0.219 | 11 | Yes |
| Spirillum minus | Bacteria | Rat Bite Fever | 1 | - | - | No |
| Sporothrix schenckii | Fungal Spore | Sporotrichosis | 6.325 | - | - | Yes |
| Staphylococcus aureus (MRSA) | Bacteria | MRSA, various infections | 0.866 | 0.596 | 4 | Yes |
| Streptobacillus moniliformis | Bacteria | Rat Bite Fever | 0.707 | - | - | No |
| Streptococcus pyogenes | Bacteria | fever | 0.894 | 1.561 | 1 | Yes |
| Swine Influenza | Virus | H1N1 flu | 0.1 | 0.098 | 23 | Yes |
| Trichophyton spp. | Fungal Spore | Dermatophytosis | 7 | 0.00411 | 560 | Yes |
| Yersinia pestis | Bacteria | Bubonic & Pneumonic Plague | 0.707 | 0.106 | 22 | Yes |

Table 4A: UV24 Removal Rates for Cat Pathogens

| PATHOGEN | Mean | MERV 6 | UV k | Single Pass | Total Removal |
|----------------------------------------------|--------------------|---------------|-----------------------|-----------------|---------------|
| | Dia. μm | Filtration, % | m^2/J | UV Kill Rate, % | Rate, % |
| Aspergillus spp. | 3.354 | 44.76 | 0.00058 | 10.85 | 50.75 |
| Avian Influenza A | 0.098 | 7.09 | 0.101 | 100.00 | 100.00 |
| Bacillus anthracis | 1.118 | 18.79 | 0.02702 | 99.53 | 99.61 |
| Bordetella bronchiseptica | 0.707 | 10.57 | 0.0364 | 99.93 | 99.93 |
| Brachyspira pilosicoli | 0.5 | 6.89 | 0.0324 | 99.84 | 99.85 |
| Burkholderia mallei | 0.77 | 11.80 | 0.034 | 99.88 | 99.89 |
| Burkholderia pseudomallei | 0.494 | 6.80 | 0.0344 | 99.89 | 99.90 |
| Campylobacter coli | 2.12 | 35.03 | 0.144 | 100.00 | 100.00 |
| Campylobacter jejuni | 2.12 | 35.03 | 0.1444 | 100.00 | 100.00 |
| Canine Distemper Virus (CDV) | 0.15 | 5.09 | 0.11 | 100.00 | 100.00 |
| Canine Influenza H3N2 | 0.098 | 7.09 | 0.101 | 100.00 | 100.00 |
| Canine Parvovirus 2 | 0.022 | 21.04 | 0.092 | 100.00 | 100.00 |
| Chlamydomphila felis | 0.283 | 4.42 | 0.0384 | 99.95 | 99.95 |
| Clostridium botulinum | 1.975 | 33.20 | 0.0412 | 99.97 | 99.98 |
| Clostridium tetani | 5 | 48.83 | 0.04699 | 99.99 | 100.00 |
| Coccidioides immitis | 3.464 | 45.25 | - | - | - |
| Cowpox | 0.173 | 4.69 | 0.135 | 100.00 | 100.00 |
| Coxiella burnettii | 0.283 | 4.42 | 0.1535 | 100.00 | 100.00 |
| Cryptococcus neoformans | 4.899 | 48.72 | 0.0167 | 96.34 | 98.12 |
| Epidermophyton | 8.5 | 49.96 | - | - | - |
| Erysipelothrix spp. | 1 | 16.43 | - | - | - |
| Feline Calicivirus (FeCV) | 0.037 | 15.49 | 0.0345 | 99.89 | 99.91 |
| Feline Distemper (Feline Parvovirus, FPV) | 0.022 | 21.04 | 0.092 | 100.00 | 100.00 |
| Feline Herpesvirus T1 (Rhinotracheitis, FVR) | 0.18 | 4.60 | 0.1046 | 100.00 | 100.00 |
| Feline Infectious Peritonitis virus (FIP) | 0.11 | 6.43 | 0.0053 | 64.99 | 67.24 |
| Feline Influenza A (H7N2) | 0.098 | 7.09 | 0.101 | 100.00 | 100.00 |
| Feline Panleukopenia (Picornavirus) (FPV) | 0.037 | 15.49 | 0.066 | 100.00 | 100.00 |
| Francisella tularensis | 0.2 | 4.43 | 0.009 | 83.17 | 83.91 |
| Hantavirus (Hantaan Virus) | 0.095 | 7.28 | 0.0688 | 100.00 | 100.00 |
| Hendra Virus | 0.175 | 4.67 | 0.23026 | 100.00 | 100.00 |
| Histoplasma capsulatum | 2.236 | 36.37 | 0.01645 | 96.15 | 97.55 |
| Influenza A virus | 0.098 | 7.09 | 0.101 | 100.00 | 100.00 |
| Leptospira spp. | 0.1 | 6.97 | - | - | - |
| Listeria monocytogenes | 0.707 | 10.57 | 0.0127 | 91.91 | 92.77 |
| Microsporium spp. | 2.96 | 42.60 | - | - | - |
| Mycobacterium avium | 1.118 | 18.79 | 0.04387 | 99.98 | 99.99 |
| Mycobacterium bovis | 0.637 | 9.25 | 0.181 | 100.00 | 100.00 |
| Mycobacterium tuberculosis | 0.637 | 9.25 | 0.472 | 100.00 | 100.00 |
| Mycoplasma spp. | 0.177 | 4.64 | 0.284 | 100.00 | 100.00 |
| Nipah virus (Henipah virus) | 0.175 | 4.67 | 0.32894 | 100.00 | 100.00 |
| Pneumocystis carinii | 2 | 33.53 | - | - | - |
| Pseudorabies (PRV) | 0.194 | 4.47 | 0.0676 | 100.00 | 100.00 |
| Rabies virus | 0.07 | 9.46 | 0.219 | 100.00 | 100.00 |
| Spirillus minus | 1 | 16.43 | - | - | - |
| Sporothrix schenckii | 6.325 | 49.66 | - | - | - |
| Staphylococcus aureus (MRSA) | 0.866 | 13.72 | 0.596 | 100.00 | 100.00 |
| Streptobacillus moniliformis | 0.707 | 10.57 | - | - | - |
| Streptococcus pyogenes | 0.894 | 14.28 | 1.561 | 100.00 | 100.00 |
| Swine Influenza | 0.1 | 6.97 | 0.098 | 100.00 | 100.00 |
| Trichophyton spp. | 7 | 49.82 | 0.00411 | 55.68 | 77.76 |
| Yersinia pestis | 0.707 | 10.57 | 0.106 | 100.00 | 100.00 |

Table 4A summarizes the calculated removal rates for all the pathogens in Table 4, except where the UV rate constant is unknown. Figure 4 illustrates the removal rates of the pathogens in Table 4A in a model room of 800 ft³ with no outside air and this graph only includes those pathogens with known UV rate constants. It can be observed in Figure 4 that all pathogens are removed to near-zero levels within about one hour.

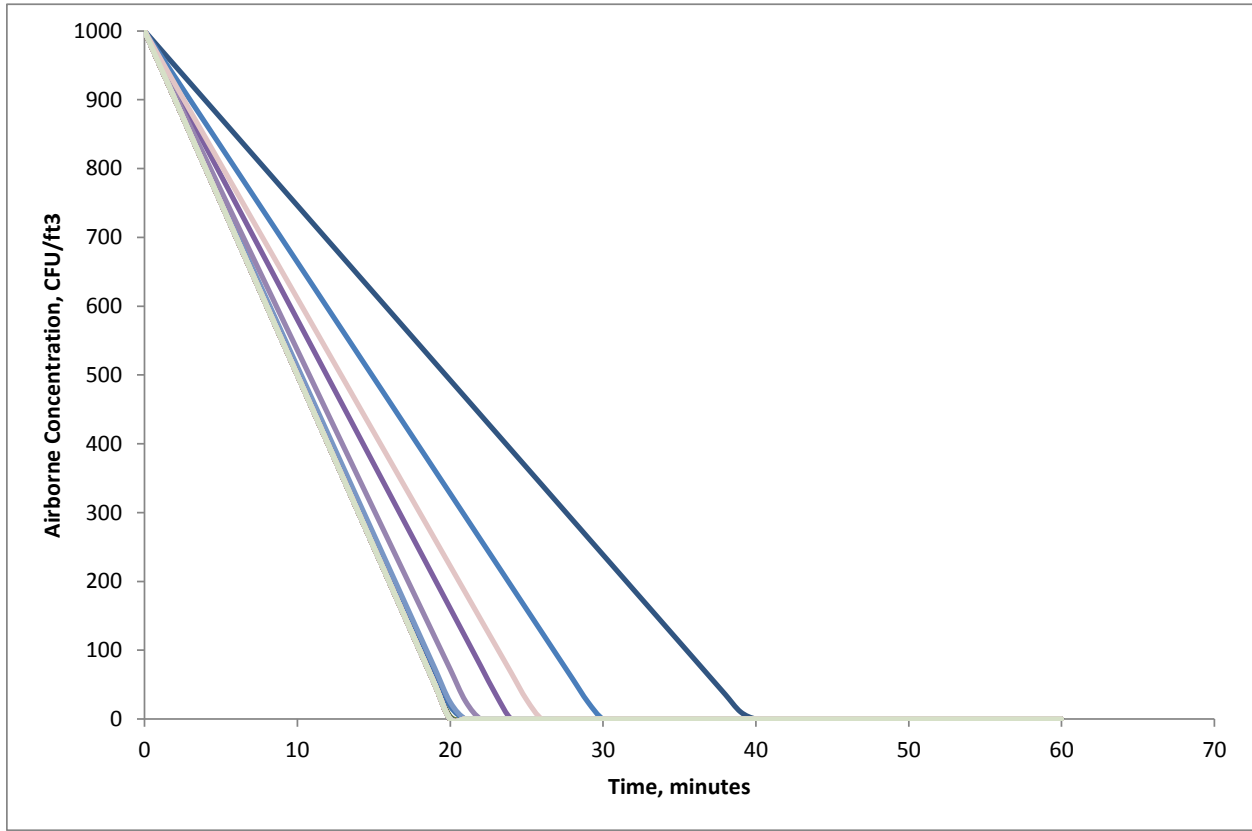


Figure 4: Removal of Feline pathogens by the UV24 unit in a room of 800 ft³ volume with no outside air. Only microbes from Table 4A with known UV rate constants are included.

Airborne Diseases of Birds and Poultry

Table 5 lists all of the potentially airborne zoonotic pathogens of birds (pet birds) and poultry. Table 5A summarizes the calculated removal rates for all the pathogens in Table 5, except where the UV rate constant is unknown. Figure 5 illustrates the removal rates of the pathogens in Table 4A in a model room of 800 ft³ with no outside air and this graph only includes those pathogens with known UV rate constants. It can be observed in Figure 5 that all pathogens are removed to near-zero levels within about one hour.

Table 5: Potentially Airborne Bird & Poultry Pathogens and Allergens

| <u>PATHOGEN or DISEASE</u> | <u>GROUP</u> | <u>DISEASE</u> | <u>Mean</u> | <u>UV k</u> | <u>UV D90</u> | <u>Trans to</u> |
|-------------------------------------------------|---------------------|-----------------------------------|---------------------------------------------|------------------------------------------------|------------------------------------------------|------------------------|
| | | | <u>dia. μm</u> | <u>m^2/J</u> | <u>J/m^2</u> | <u>Human</u> |
| Aspergillus spp. | Fungal Spore | Aspergillosis | 3.354 | 0.00058 | 3970 | Yes |
| Avian Adenovirus (FAV) | Virus | respiratory disease, bronchitis | 0.08 | 0.0108 | 213 | No |
| Avian Encephalomyelitis Virus | Virus | Encephalomyelitis | 0.023 | 0.02303 | 100 | No |
| Avian Influenza A | Virus | flu | 0.098 | 0.101 | 23 | Yes |
| Avian Leukosis virus (RSA) | Virus | leukosis | 0.107 | 0.00365 | 631 | No |
| Avian metapneumovirus | Virus | turkey rhinotracheitis | 0.1 | - | - | No |
| Avian Sarcoma virus | Virus | sarcoma | 0.098 | 0.0105 | 219 | No |
| Bordetella avium | Bacteria | Bordetellosis | 0.7 | 0.3838 | 6 | No |
| Campylobacter jejuni | Bacteria | Enteritis | 2.12 | 0.1444 | 16 | Yes |
| Canary Pox Virus | Virus | pox | 0.24 | 0.1439 | 16 | No |
| Candida | Fungi | Candidiasis | 4.899 | 0.01 | 230 | Yes |
| Chlamydia caviae (formerly C.psittaci) | Bacteria | Psittacosis, parrot fever | 0.283 | 0.039 | 59 | Yes |
| Clostridium botulinum | Bacteria | Botulism | 1.975 | 0.0412 | 56 | Yes |
| Clostridium perfringens | Bacteria | sepsis, toxins, food poisoning | 5 | 0.06 | 38 | Yes |
| Coronavirus | Virus | Infectious bronchitis | 0.113 | 0.01 | 230 | Yes |
| Cryptococcus neoformans | Fungi | Cryptococcosis | 4.899 | 0.0167 | 138 | Yes |
| Feline Influenza A (H7N2) | Virus | Flu | 0.10 | 0.101 | 23 | Yes |
| Fowlpox virus (Avian poxvirus) | Virus | fowlpox | 0.24 | 0.1396 | 16 | Yes |
| Histoplasma capsulatum | Fungal Spore | URD | 2.236 | 0.01645 | 140 | Yes |
| Infectious Bronchitis Virus (IBV) | Virus | bronchitis | 0.113 | 0.2424 | 9 | No |
| Infectious Laryngotracheitis (Psittacid herpes) | Virus | respiratory disease | 0.18 | 0.1151 | 20 | No |
| Influenza A virus | Virus | flu, secondary pneumonia | 0.098 | 0.101 | 23 | Yes |
| Marek's Disease Virus (Herpesvirus) | Virus | Marek's disease, LT | 0.18 | 0.06325 | 36 | No |
| Mycobacterium avium | Bacteria | Paratuberculosis, Johne's Disease | 1.118 | 0.04387 | 52 | Yes |
| Mycoplasma gallisepticum | Bacteria | respiratory disease | 0.177 | 0.284 | 8 | No |
| Mycoplasma spp. | Bacteria | infectious anemia, eye disease | 0.177 | 0.284 | 8 | Yes |
| Mycoplasma synoviae | Bacteria | Synovitis | 0.177 | 0.284 | 8 | No |
| Newcastle Disease Virus (NDV) | Virus | NDV | 0.212 | 0.1636 | 14 | Yes |
| Ornithobacterium rhinotracheale (ORT) | Bacteria | bacterial infection | 0.64 | - | - | No |
| Papilloma virus | Virus | wart-like tumors | 0.055 | 0.0256 | 90 | No |
| Polyomavirus | Virus | paralysis, diarrhea | 0.0424 | 0.00408 | 564 | No |
| Pseudomonas aeruginosa | Bacteria | infection | 0.494 | 0.128 | 18 | Yes |
| Psittacine Beak and Feather Disease (Pbfd) | Virus | beak & feather infections | 0.055 | 0.007 | 329 | No |
| Reovirus | Virus | colds, fever, pneumonia | 0.08 | 0.016 | 144 | Yes |
| Salmonella enteritidis | Bacteria | Salmonellosis | 0.81 | 0.221 | 10 | Yes |
| Salmonella typhi | Bacteria | Salmonellosis | 0.81 | 0.1467 | 16 | Yes |
| Staphylococcus aureus (MRSA) | Bacteria | MRSA, various infections | 0.866 | 0.596 | 4 | Yes |

Table 5A: UV24 Removal Rates for Bird Pathogens

| PATHOGEN | Mean | MERV 6 | UV k | Single Pass | Total Removal |
|-------------------------------------------------|--------------------|---------------|-----------------------|-----------------|---------------|
| | Dia. μm | Filtration, % | m^2/J | UV Kill Rate, % | Rate, % |
| Aspergillus spp. | 3.354 | 44.76 | 0.00058 | 10.85 | 50.75 |
| Avian Adenovirus (FAV) | 0.08 | 8.44 | 0.0108 | 88.22 | 89.21 |
| Avian Encephalomyelitis Virus | 0.023 | 20.57 | 0.02303 | 98.95 | 99.17 |
| Avian Influenza A | 0.098 | 7.09 | 0.101 | 100.00 | 100.00 |
| Avian Leukosis virus (RSA) | 0.107 | 6.58 | 0.00365 | 51.46 | 54.65 |
| Avian metapneumovirus | 0.1 | 6.97 | - | - | - |
| Avian Sarcoma virus | 0.098 | 7.09 | 0.0105 | 87.49 | 88.38 |
| Bordetella avium | 0.7 | 10.44 | 0.3838 | 100.00 | 100.00 |
| Campylobacter jejuni | 2.12 | 35.03 | 0.1444 | 100.00 | 100.00 |
| Canary Pox Virus | 0.24 | 4.31 | 0.1439 | 100.00 | 100.00 |
| Candida | 4.899 | 48.72 | 0.01 | 86.19 | 92.92 |
| Chlamydophila caviae (formerly C.psittaci) | 0.283 | 4.42 | 0.039 | 99.96 | 99.96 |
| Clostridium botulinum | 1.975 | 33.20 | 0.0412 | 99.97 | 99.98 |
| Clostridium perfringens | 5 | 48.83 | 0.06 | 100.00 | 100.00 |
| Coronavirus | 0.113 | 6.29 | 0.01 | 86.19 | 87.06 |
| Cryptococcus neoformans | 4.899 | 48.72 | 0.0167 | 96.34 | 98.12 |
| Feline Influenza A (H7N2) | 0.098 | 7.09 | 0.101 | 100.00 | 100.00 |
| Fowlpox virus (Avian poxvirus) | 0.24 | 4.31 | 0.1396 | 100.00 | 100.00 |
| Histoplasma capsulatum | 2.236 | 36.37 | 0.01645 | 96.15 | 97.55 |
| Infectious Bronchitis Virus (IBV) | 0.113 | 6.29 | 0.2424 | 100.00 | 100.00 |
| Infectious Laryngotracheitis (Psittacid herpes) | 0.18 | 4.60 | 0.1151 | 100.00 | 100.00 |
| Influenza A virus | 0.098 | 7.09 | 0.101 | 100.00 | 100.00 |
| Marek's Disease Virus (Herpesvirus) | 0.18 | 4.60 | 0.06325 | 100.00 | 100.00 |
| Mycobacterium avium | 1.118 | 18.79 | 0.04387 | 99.98 | 99.99 |
| Mycoplasma gallisepticum | 0.177 | 4.64 | 0.284 | 100.00 | 100.00 |
| Mycoplasma spp. | 0.177 | 4.64 | 0.284 | 100.00 | 100.00 |
| Mycoplasma synoviae | 0.177 | 4.64 | 0.284 | 100.00 | 100.00 |
| Newcastle Disease Virus (NDV) | 0.212 | 4.36 | 0.1636 | 100.00 | 100.00 |
| Ornithobacterium rhinotracheale (ORT) | 0.64 | 9.31 | - | - | - |
| Papilloma virus | 0.055 | 11.55 | 0.0256 | 99.37 | 99.44 |
| Polyomavirus | 0.0424 | 14.08 | 0.00408 | 55.42 | 61.70 |
| Pseudomonas aeruginosa | 0.494 | 6.80 | 0.128 | 100.00 | 100.00 |
| Psittacine Beak and Feather Disease (PBFD) | 0.055 | 11.55 | 0.007 | 74.99 | 77.88 |
| Reovirus | 0.08 | 8.44 | 0.016 | 95.79 | 96.15 |
| Salmonella enteritidis | 0.81 | 12.59 | 0.221 | 100.00 | 100.00 |
| Salmonella typhi | 0.81 | 12.59 | 0.1467 | 100.00 | 100.00 |
| Staphylococcus aureus (MRSA) | 0.866 | 13.72 | 0.596 | 100.00 | 100.00 |

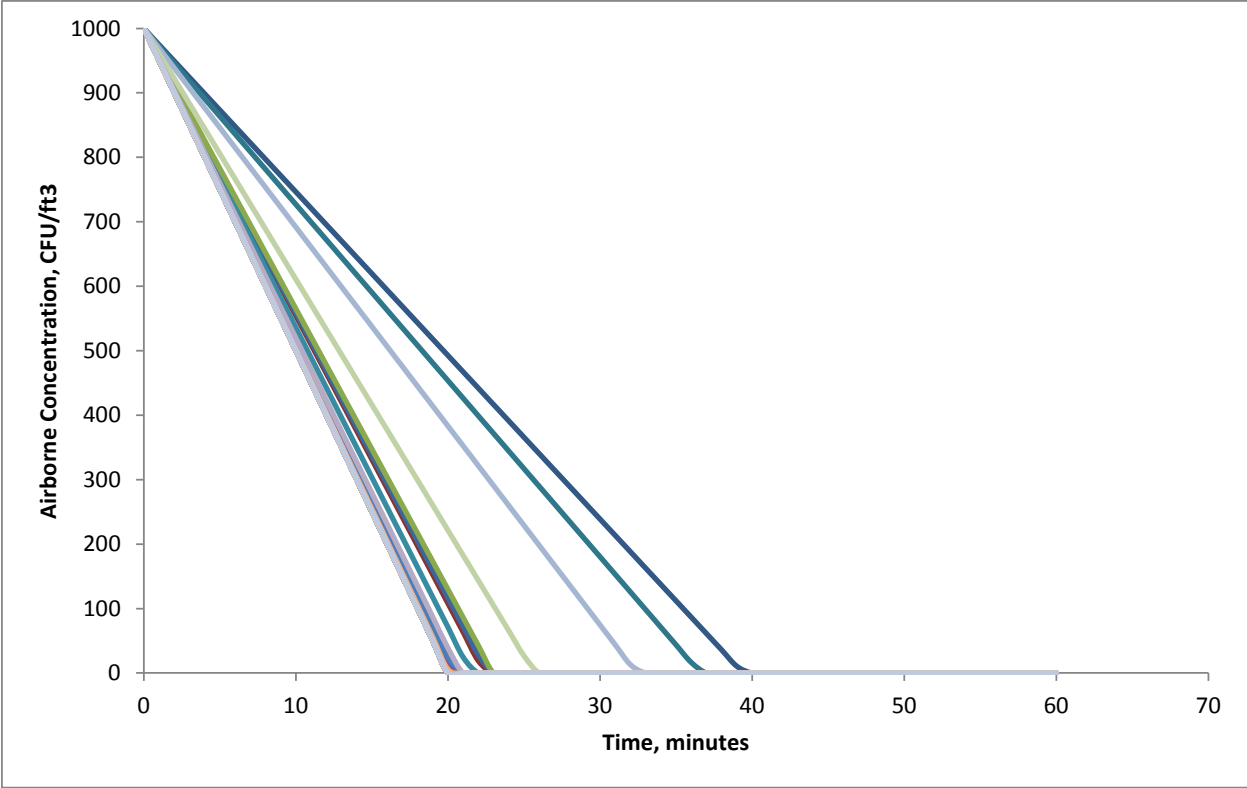


Figure 5: Removal of Bird pathogens by the UV24 unit in a room of 800 ft³ volume with no outside air. Only microbes from Table 5A with known UV rate constants are included.

Airborne Diseases of Rodents, Hamsters, and Rabbits

Table 6 lists all of the potentially airborne zoonotic pathogens of rodents, hamsters and rabbits. Table 6A summarizes the calculated removal rates for all the pathogens in Table 6, except where the UV rate constant is unknown. Figure 6 illustrates the removal rates of the pathogens in Table 6A in a model room of 800 ft³ with no outside air and this graph only includes those pathogens with known UV rate constants. It can be observed in Figure 6 that all pathogens are removed to near-zero levels within about one hour.

Table 6: Potentially Airborne Pathogens and Allergens of Rodents, Hamsters and Rabbits

| PATHOGEN or DISEASE | GROUP | DISEASE | Mean | UV k | UV D90 | Trans to Human |
|--------------------------------------------|-----------------|-----------------------------------|--------------------|-----------------------|-----------------------|----------------|
| | | | Dia. μm | m^2/J | J/m^2 | |
| Actinomyces bovis | Bacteria | actinomycosis | 0.5 | - | - | No |
| Actinomyces israeli | Bacteria | actinomycosis | 0.5 | - | - | No |
| Aerococcus viridans | Bacteria | meningitis | 1 | - | - | No |
| Aeromonas spp. | Bacteria | bacteremia | 2.098 | 0.2031 | 11 | Yes |
| Bacillus anthracis | Bacterial Spore | Anthrax | 1.118 | 0.02702 | 85 | Yes |
| Bacteroides fragilis | Bacteria | opportunistic infection | 3.162 | 0.0675 | 34 | Yes |
| Bordetella bronchiseptica | Bacteria | kennel cough | 0.707 | 0.0364 | 63 | Yes |
| Burkholderia mallei | Bacteria | Glanders | 0.77 | 0.034 | 68 | Yes |
| Burkholderia pseudomallei | Bacteria | Melioidosis | 0.494 | 0.0344 | 67 | Yes |
| Chlamydomydia muridarum | Bacteria | Chlamydiosis | 0.283 | - | - | No |
| Chlamydomydia caviae (formerly C.psittaci) | Bacteria | Psittacosis, parrot fever | 0.283 | 0.039 | 59 | Yes |
| Clostridium tetani | Bacteria | tetanus | 5 | 0.04699 | 49 | Yes |
| Coccidioides immitis | Fungal Spore | Coccidioidomycosis | 3.464 | - | - | Yes |
| Coxiella burnetii | Bacteria | Q Fever | 0.283 | 0.1535 | 15 | Yes |
| Coxsackievirus | Virus | colds | 0.027 | 0.02834 | 81 | Yes |
| Echovirus | Virus | colds | 0.024 | 0.02786 | 83 | Yes |
| Francisella tularensis | Bacteria | tularemia, pneumonia, fever | 0.2 | 0.009 | 256 | Yes |
| Guineapig adenovirus | Virus | colds | 0.079 | 0.0026 | 886 | - |
| Hantavirus (Hantaan Virus) | Virus | Hemorrhagic Fever | 0.095 | 0.0688 | 33 | Yes |
| Haemophilus spp. | Bacteria | pneumonia, conjunctivitis | 0.285 | 0.0599 | 38 | Yes |
| Influenza A virus | Virus | flu, secondary pneumonia | 0.098 | 0.101 | 23 | Yes |
| Klebsiella pneumoniae | Bacteria | pneumonia | 0.671 | 0.04435 | 52 | Yes |
| Kyasanur Forest virus | Virus | viral disease | 0.045 | - | - | No |
| Leptospira spp. | Spirochete | Leptospirosis | 0.1 | NA | - | Yes |
| Listeria monocytogenes | Bacteria | Listeriosis | 0.707 | 0.0127 | 181 | Yes |
| Louping Ill (LIV) | Virus | Encephalomyelitis | 0.05 | 0.003289 | 700 | Yes |
| Lymphocytic choriomeningitis (LCMV) | Virus | Armstrong's disease | 0.087 | 0.0605 | 38 | Yes |
| Microsporum spp. | Fungal Spore | Dermatophytosis | 2.96 | - | - | Yes |
| Mumps | Virus | mumps | 0.245 | 0.077 | 30 | Yes |
| Mycobacterium avium | Bacteria | Paratuberculosis, Johne's Disease | 1.118 | 0.04387 | 52 | Yes |
| Mycobacterium bovis | Bacteria | Tuberculosis | 0.637 | 0.181 | 13 | Yes |
| Mycoplasma spp. | Bacteria | infectious anemia, eye disease | 0.177 | 0.284 | 8 | Yes |
| Myxoma virus | Virus | Myxomatosis | 0.3 | unk | unk | No |
| Nipah virus (Henipah virus) | Virus | respiratory syndrome | 0.175 | 0.328941 | 7 | No |
| Omsk hemorrhagic fever | Virus | hemorrhagic fever | 0.043 | - | - | No |
| Pasteurella multocida | Bacteria | Pasteurellosis, snuffles | 0.6 | unk | unk | Yes |
| Parainfluenza virus | Virus | flu, colds, croup, pneumonia | 0.194 | 0.1086 | 21 | Yes |
| Pseudomonas diminuta | Bacteria | rhinitis | 0.494 | 0.128 | 18 | Yes |
| Pseudorabies (PRV) | Virus | Aujeszky's Disease Virus (ADV) | 0.194 | 0.0676 | 34 | No |
| Rabies virus | Virus | rabies | 0.07 | 0.219 | 11 | Yes |
| Rabbit Haemorrhagic Disease | Virus | Rabbit Calicivirus (prev.) | 0.0307 | unk | unk | No |
| Reovirus | Virus | colds, fever, pneumonia | 0.08 | 0.016 | 144 | Yes |
| Salmonella enteritidis | Bacteria | Salmonellosis | 0.81 | 0.221 | 10 | Yes |
| Salmonella typhi | Bacteria | Salmonellosis | 0.81 | 0.1467 | 16 | Yes |
| Spirillum minus | Bacteria | Rat Bite Fever | 1 | - | - | No |
| Sporothrix schenckii | Fungal Spore | Sporotrichosis | 6.325 | - | - | Yes |
| Staphylococcus aureus (MRSA) | Bacteria | MRSA, various infections | 0.866 | 0.596 | 4 | Yes |
| Streptobacillus moniliformis | Bacteria | Rat Bite Fever | 0.707 | - | - | No |
| Streptococcus pneumoniae | Bacteria | pneumonia | 0.71 | 0.00492 | 468 | Yes |
| Streptococcus pyogenes | Bacteria | fever | 0.894 | 1.561 | 1 | Yes |
| Trichophyton spp. | Fungal Spore | Dermatophytosis | 7 | 0.00411 | 560 | Yes |
| Yersinia pestis | Bacteria | Bubonic & Pneumonic Plague | 0.707 | 0.106 | 22 | Yes |
| Yersinia pseudotuberculosis | Bacteria | pseudotuberculosis | 0.63 | 0.106 | 22 | Yes |

Table 6A: UV24 Removal Rates for Rodent Pathogens

| PATHOGEN | Mean | MERV 6 | UV k | Single Pass | Total Removal |
|--------------------------------------------|--------------------|---------------|-----------------------|-----------------|---------------|
| | Dia. μm | Filtration, % | m^2/J | UV Kill Rate, % | Rate, % |
| Actinomyces bovis | 0.5 | 6.89 | - | - | - |
| Actinomyces israeli | 0.5 | 6.89 | - | - | - |
| Aerococcus viridans | 1 | 16.43 | - | - | - |
| Aeromonas spp. | 2.098 | 34.77 | 0.2 | 100.00 | 100.00 |
| Bacillus anthracis | 1.118 | 18.79 | 0.03 | 99.53 | 99.61 |
| Bacteroides fragilis | 3.162 | 43.79 | 0.07 | 100.00 | 100.00 |
| Bordetella bronchiseptica | 0.707 | 10.57 | 0.04 | 99.93 | 99.93 |
| Burkholderia mallei | 0.77 | 11.80 | 0.03 | 99.88 | 99.89 |
| Burkholderia pseudomallei | 0.494 | 6.80 | 0.03 | 99.89 | 99.90 |
| Chlamydophila muridarum | 0.283 | 4.42 | - | - | - |
| Chlamydophila caviae (formerly C.psittaci) | 0.283 | 4.42 | 0.04 | 99.96 | 99.96 |
| Clostridium tetani | 5 | 48.83 | 0.05 | 99.99 | 100.00 |
| Coccidioides immitis | 3.464 | 45.25 | - | - | - |
| Coxiella burnettii | 0.283 | 4.42 | 0.15 | 100.00 | 100.00 |
| Coxsackievirus | 0.027 | 18.86 | 0.03 | 99.63 | 99.70 |
| Echovirus | 0.024 | 20.12 | 0.03 | 99.60 | 99.68 |
| Francisella tularensis | 0.2 | 4.43 | 0.01 | 83.17 | 83.91 |
| Guineapig adenovirus | 0.079 | 8.53 | 0 | 40.24 | 45.34 |
| Hantavirus (Hantaan Virus) | 0.095 | 7.28 | 0.07 | 100.00 | 100.00 |
| Haemophilus spp. | 0.285 | 4.43 | 0.06 | 100.00 | 100.00 |
| Influenza A virus | 0.098 | 7.09 | 0.1 | 100.00 | 100.00 |
| Klebsiella pneumoniae | 0.671 | 9.88 | 0.04 | 99.98 | 99.99 |
| Kyasanur Forest virus | 0.045 | 13.48 | - | - | - |
| Leptospira spp. | 0.1 | 6.97 | - | - | - |
| Listeria monocytogenes | 0.707 | 10.57 | 0.01 | 91.91 | 92.77 |
| Louping III (LIV) | 0.05 | 12.45 | 0 | 47.86 | 54.35 |
| Lymphocytic choriomeningitis (LCMV) | 0.087 | 7.85 | 0.06 | 100.00 | 100.00 |
| Microsporum spp. | 2.96 | 42.60 | - | - | - |
| Mumps | 0.245 | 4.31 | 0.08 | 100.00 | 100.00 |
| Mycobacterium avium | 1.118 | 18.79 | 0.04 | 99.98 | 99.99 |
| Mycobacterium bovis | 0.637 | 9.25 | 0.18 | 100.00 | 100.00 |
| Mycoplasma spp. | 0.177 | 4.64 | 0.28 | 100.00 | 100.00 |
| Myxoma virus | 0.3 | 4.52 | unk | - | - |
| Nipah virus (Henipah virus) | 0.175 | 4.67 | 0.33 | 100.00 | 100.00 |
| Omsk hemorrhagic fever | 0.043 | 13.94 | - | - | - |
| Pasteurella multocida | 0.6 | 8.58 | unk | - | - |
| Parainfluenza virus | 0.194 | 4.47 | 0.11 | 100.00 | 100.00 |
| Pseudomonas diminuta | 0.494 | 6.80 | 0.13 | 100.00 | 100.00 |
| Pseudorabies (PRV) | 0.194 | 4.47 | 0.07 | 100.00 | 100.00 |
| Rabies virus | 0.07 | 9.46 | 0.22 | 100.00 | 100.00 |
| Rabbit Haemorrhagic Disease | 0.0307 | 17.48 | unk | - | - |
| Reovirus | 0.08 | 8.44 | 0.02 | 95.79 | 96.15 |
| Salmonella enteritidis | 0.81 | 12.59 | 0.22 | 100.00 | 100.00 |
| Salmonella typhi | 0.81 | 12.59 | 0.15 | 100.00 | 100.00 |
| Spirillum minus | 1 | 16.43 | - | - | - |
| Sporothrix schenckii | 6.325 | 49.66 | - | - | - |
| Staphylococcus aureus (MRSA) | 0.866 | 13.72 | 0.6 | 100.00 | 100.00 |
| Streptobacillus moniliformis | 0.707 | 10.57 | - | - | - |
| Streptococcus pneumoniae | 0.71 | 10.63 | 0 | 62.25 | 66.26 |
| Streptococcus pyogenes | 0.894 | 14.28 | 1.56 | 100.00 | 100.00 |
| Trichophyton spp. | 7 | 49.82 | 0 | 55.68 | 77.76 |
| Yersinia pestis | 0.707 | 10.57 | 0.11 | 100.00 | 100.00 |
| Yersinia pseudotuberculosis | 0.63 | 9.12 | 0.11 | 100.00 | 100.00 |

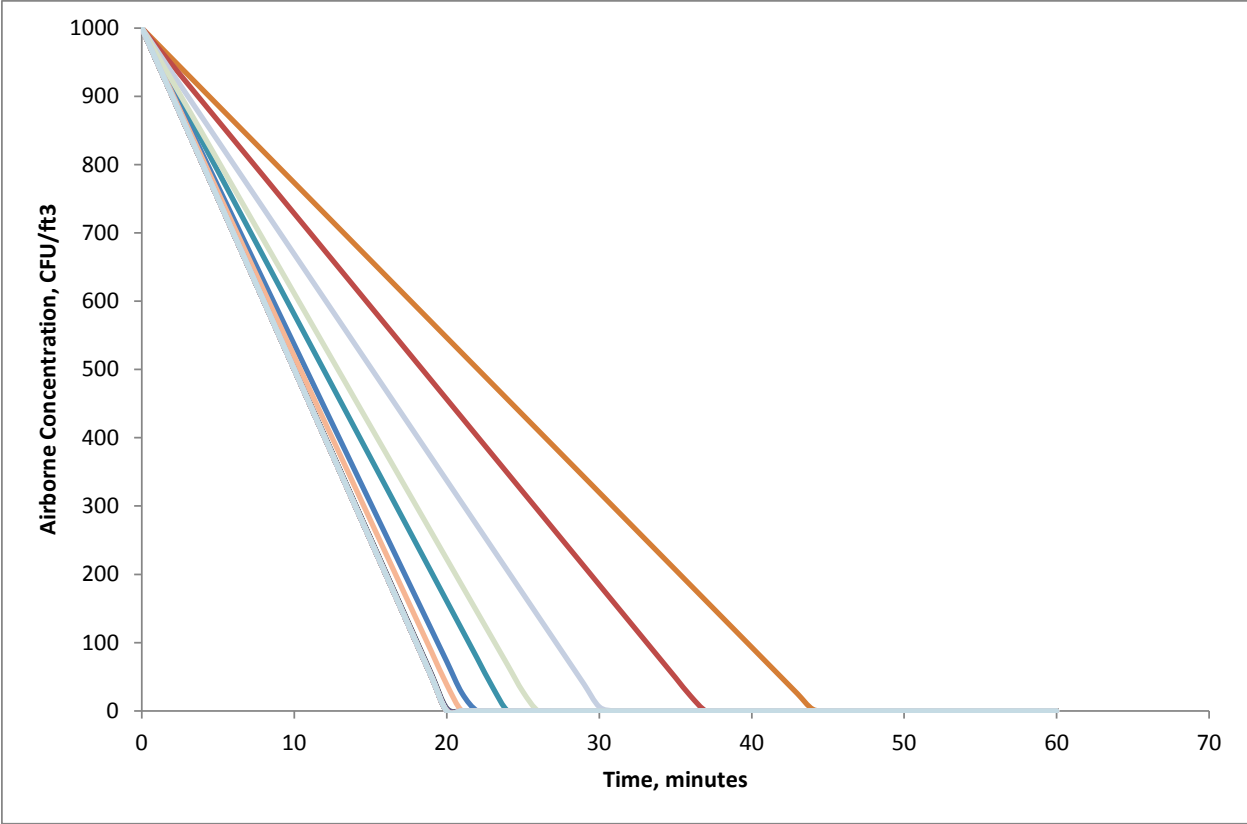


Figure 6: Removal of Rodent pathogens by the UV24 unit in a room of 800 ft³ volume with no outside air. Only microbes from Table 6A with known UV rate constants are included.

Airborne Diseases of Horses

Table 7 lists all of the potentially airborne zoonotic pathogens of horses. Table 7A summarizes the calculated removal rates for all the pathogens in Table 6, except where the UV rate constant is unknown. Figure 7 illustrates the removal rates of the pathogens in Table 7A in a model room of 800 ft³ with no outside air and this graph only includes those pathogens with known UV rate constants. It can be observed in Figure 7 that all pathogens are removed to near-zero levels within about one hour.

Table 7: Potentially Airborne Pathogens and Allergens of Horses

| PATHOGEN or DISEASE | GROUP | DISEASE | Mean Dia. μm | UV k m^2/J | UV D90 J/m^2 | Trans to Human |
|------------------------------|-----------------|-----------------------------------|-----------------------------------------------|--------------------------------------------------|----------------------------------------------------|---------------------------|
| Arterivirus | Virus | EAV, PRRD | 0.05 | - | - | No |
| Avian Influenza A | Virus | Flu | 0.098 | 0.101 | 23 | Yes |
| Bacillus anthracis | Bacterial Spore | Anthrax | 1.118 | 0.02702 | 85 | Yes |
| Brucella abortus | Bacteria | Brucellosis, undulant fever | 0.57 | 0.0307 | 75 | Yes |
| Brucella canis | Bacteria | Brucellosis, undulant fever | 0.566 | 0.048 | 48 | Yes |
| Brucella maris | Bacteria | Brucellosis | 0.566 | 0.047 | 49 | No |
| Brucella melitensis | Bacteria | Brucellosis | 0.566 | 0.047 | 49 | Yes |
| Brucella neotomae | Bacteria | Brucellosis | 0.566 | 0.047 | 49 | No |
| Brucella ovis | Bacteria | Brucellosis | 0.566 | 0.047 | 49 | No |
| Brucella suis | Bacteria | Brucellosis | 0.57 | 0.047 | 49 | Yes |
| Burkholderia mallei | Bacteria | Glanders | 0.77 | 0.034 | 68 | Yes |
| Burkholderia pseudomallei | Bacteria | Melioidosis | 0.494 | 0.0344 | 67 | Yes |
| Canine Influenza H3N8 | Virus | Flu | 0.098 | 0.101 | 23 | No |
| Clostridium botulinum | Bacteria | Botulism | 1.975 | 0.0412 | 56 | Yes |
| Clostridium difficile | Bacteria | diarrhea | 0.6 | 0.0385 | 60 | Yes |
| Clostridium perfringens | Bacteria | sepsis, toxins, food poisoning | 5 | 0.06 | 38 | Yes |
| Clostridium tetani | Bacteria | tetanus | 5 | 0.04699 | 49 | Yes |
| Coccidioides immitis | Fungal Spore | Coccidioidomycosis | 3.464 | - | - | Yes |
| Coxiella burnetii | Bacteria | Q Fever | 0.283 | 0.1535 | 15 | Yes |
| Cryptococcus farciminosus | Fungi | Cryptococcosis | 4.9 | 0.0167 | 138 | Yes |
| Cryptococcus neoformans | Fungi | Cryptococcosis | 4.899 | 0.0167 | 138 | Yes |
| Dermatophilus congolensis | Bacteria | dermatophilosis, mud fever | 1 | - | - | No |
| Enterobacter | Bacteria | Bacteremia | 1.414 | 0.036 | 64 | Yes |
| Epidermophyton | Fungi | Dermatophytosis | 8.5 | - | - | Yes |
| Equine Influenza | Virus | Flu | 0.1 | 0.101 | 23 | No |
| Feline Influenza A (H7N2) | Virus | Flu | 0.10 | 0.101 | 23 | Yes |
| Hendra Virus | Virus | Pneumonia | 0.175 | 0.230259 | 10 | Yes |
| Histoplasma farciminosum | Fungal Spore | URD | 2.236 | 0.01645 | 140 | No |
| Louping Ill (LIV) | Virus | Encephalomyelitis | 0.05 | 0.003289 | 700 | Yes |
| Microsporium spp. | Fungal Spore | Dermatophyopsis | 2.96 | - | - | Yes |
| Mycobacterium avium | Bacteria | Paratuberculosis, Johne's Disease | 1.118 | 0.04387 | 52 | Yes |
| Nipah virus (Henipah virus) | Virus | respiratory syndrome | 0.175 | 0.328941 | 7 | No |
| Salmonella enteritidis | Bacteria | Salmonellosis | 0.81 | 0.221 | 10 | Yes |
| Salmonella typhi | Bacteria | Salmonellosis | 0.81 | 0.1467 | 16 | Yes |
| Sporothrix schenckii | Fungal Spore | Sporotrichosis | 6.325 | - | - | Yes |
| Staphylococcus aureus (MRSA) | Bacteria | MRSA, various infections | 0.866 | 0.596 | 4 | Yes |
| Streptococcus pyogenes | Bacteria | fever | 0.894 | 1.561 | 1 | Yes |
| Trichophyton spp. | Fungal Spore | Dermatophytosis | 7 | 0.00411 | 560 | Yes |

Table 7A: UV24 Removal Rates for Horse Pathogens

| PATHOGEN | Mean | MERV 6 | UV k | Single Pass | Total Removal |
|------------------------------|--------------------|---------------|-----------------------|-----------------|---------------|
| | Dia. μm | Filtration, % | m^2/J | UV Kill Rate, % | Rate, % |
| Arterivirus | 0.05 | 12.45 | - | - | - |
| Avian Influenza A | 0.098 | 7.09 | 0.1010 | 100.00 | 100.00 |
| Bacillus anthracis | 1.118 | 18.79 | 0.0270 | 99.53 | 99.61 |
| Brucella abortus | 0.57 | 8.05 | 0.0307 | 99.77 | 99.79 |
| Brucella canis | 0.566 | 7.98 | 0.0480 | 99.99 | 99.99 |
| Brucella maris | 0.566 | 7.98 | 0.0470 | 99.99 | 99.99 |
| Brucella melitensis | 0.566 | 7.98 | 0.0470 | 99.99 | 99.99 |
| Brucella neotomae | 0.566 | 7.98 | 0.0470 | 99.99 | 99.99 |
| Brucella ovis | 0.566 | 7.98 | 0.0470 | 99.99 | 99.99 |
| Brucella suis | 0.57 | 8.05 | 0.0470 | 99.99 | 99.99 |
| Burkholderia mallei | 0.77 | 11.80 | 0.0340 | 99.88 | 99.89 |
| Burkholderia pseudomallei | 0.494 | 6.80 | 0.0344 | 99.89 | 99.90 |
| Canine Influenza H3N8 | 0.098 | 7.09 | 0.1010 | 100.00 | 100.00 |
| Clostridium botulinum | 1.975 | 33.20 | 0.0412 | 99.97 | 99.98 |
| Clostridium difficile | 0.6 | 8.58 | 0.0385 | 99.95 | 99.96 |
| Clostridium perfringens | 5 | 48.83 | 0.0600 | 100.00 | 100.00 |
| Clostridium tetani | 5 | 48.83 | 0.0470 | 99.99 | 100.00 |
| Coccidioides immitis | 3.464 | 45.25 | - | - | - |
| Coxiella burnettii | 0.283 | 4.42 | 0.1535 | 100.00 | 100.00 |
| Cryptococcus farciminosus | 4.9 | 48.72 | 0.0167 | 96.34 | 98.12 |
| Cryptococcus neoformans | 4.899 | 48.72 | 0.0167 | 96.34 | 98.12 |
| Dermatophilus congolensis | 1 | 16.43 | - | - | - |
| Enterobacter | 1.414 | 24.42 | 0.0360 | 99.92 | 99.94 |
| Epidermophyton | 8.5 | 49.96 | - | - | - |
| Equine Influenza | 0.1 | 6.97 | 0.1010 | 100.00 | 100.00 |
| Feline Influenza A (H7N2) | 0.098 | 7.09 | 0.1010 | 100.00 | 100.00 |
| Hendra Virus | 0.175 | 4.67 | 0.2303 | 100.00 | 100.00 |
| Histoplasma farciminosum | 2.236 | 36.37 | 0.0165 | 96.15 | 97.55 |
| Louping Ill (LIV) | 0.05 | 12.45 | 0.0033 | 47.86 | 54.35 |
| Microsporium spp. | 2.96 | 42.60 | - | - | - |
| Mycobacterium avium | 1.118 | 18.79 | 0.0439 | 99.98 | 99.99 |
| Nipah virus (Henipah virus) | 0.175 | 4.67 | 0.3289 | 100.00 | 100.00 |
| Salmonella enteritidis | 0.81 | 12.59 | 0.2210 | 100.00 | 100.00 |
| Salmonella typhi | 0.81 | 12.59 | 0.1467 | 100.00 | 100.00 |
| Sporothrix schenckii | 6.325 | 49.66 | - | - | - |
| Staphylococcus aureus (MRSA) | 0.866 | 13.72 | 0.5960 | 100.00 | 100.00 |
| Streptococcus pyogenes | 0.894 | 14.28 | 1.5610 | 100.00 | 100.00 |
| Trichophyton spp. | 7 | 49.82 | 0.0041 | 55.68 | 77.76 |

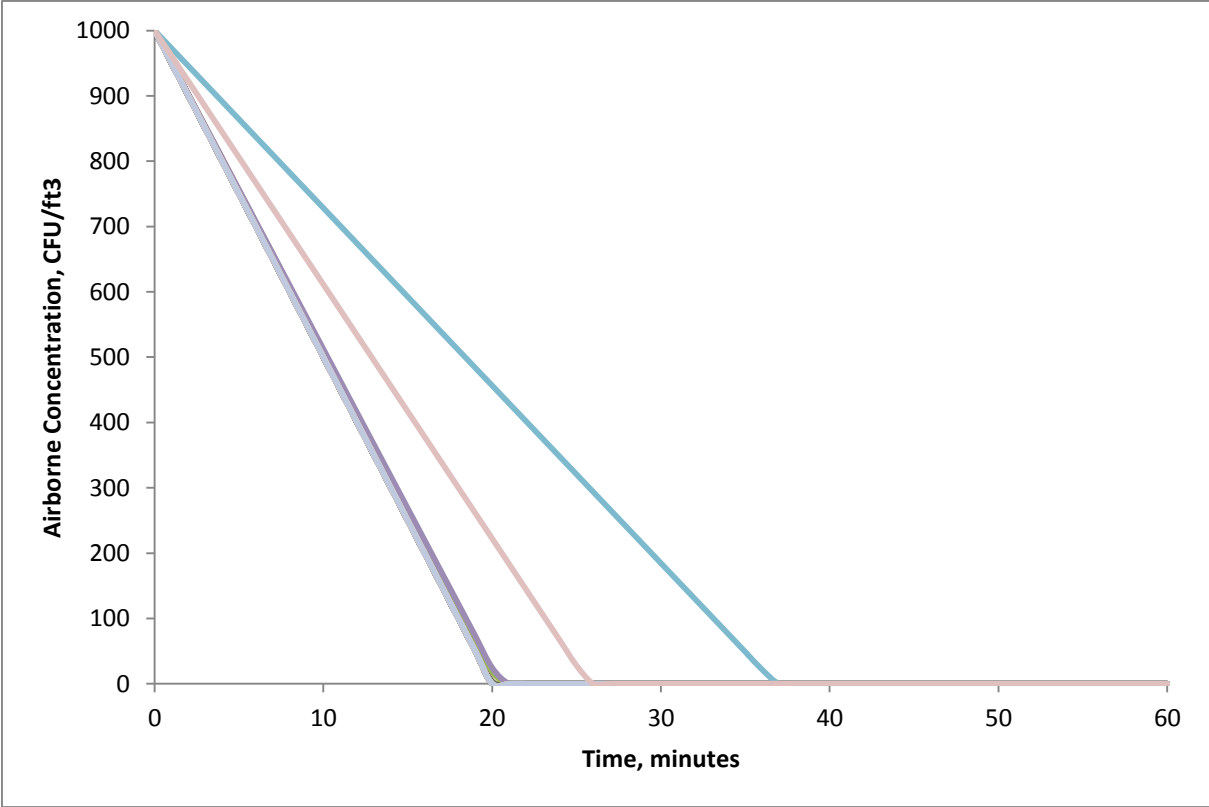


Figure 7: Removal of Horse pathogens by the UV24 unit in a room of 800 ft³ volume with no outside air. Only microbes from Table 7A with known UV rate constants are included.

Airborne Diseases of Cows

Table 8 lists all of the potentially airborne zoonotic pathogens of cows. Table 8A summarizes the calculated removal rates for all the pathogens in Table 8, except where the UV rate constant is unknown. Figure 8 illustrates the removal rates of the pathogens in Table 8A in a model room of 800 ft³ with no outside air and this graph only includes those pathogens with known UV rate constants. It can be observed in Figure 8 that all pathogens are removed to near-zero levels within about one hour.

Table 8: Potentially Airborne Pathogens and Allergens of Cows

| PATHOGEN or DISEASE | GROUP | DISEASE | Mean dia. μm | UV k m^2/J | UV D90 J/m^2 | Trans to Human |
|------------------------------|-----------------|-----------------------------------|-----------------------------------------------|--------------------------------------------------|----------------------------------------------------|---------------------------|
| Actinomyces bovis | Bacteria | actinomycosis | 0.5 | - | - | No |
| Bacillus anthracis | Bacterial Spore | Anthrax | 1.118 | 0.02702 | 85 | Yes |
| Bovine Coronavirus (BCV) | Virus | respiratory infections | 0.113 | 0.377 | 6 | No |
| Brucella abortus | Bacteria | Brucellosis, undulant fever | 0.57 | 0.0307 | 75 | Yes |
| Brucella canis | Bacteria | Brucellosis, undulant fever | 0.566 | 0.048 | 48 | Yes |
| Brucella maris | Bacteria | Brucellosis | 0.566 | 0.047 | 49 | No |
| Brucella melitensis | Bacteria | Brucellosis | 0.566 | 0.047 | 49 | Yes |
| Brucella neotomae | Bacteria | Brucellosis | 0.566 | 0.047 | 49 | No |
| Brucella ovis | Bacteria | Brucellosis | 0.566 | 0.047 | 49 | No |
| Brucella suis | Bacteria | Brucellosis | 0.57 | 0.047 | 49 | Yes |
| Burkholderia pseudomallei | Bacteria | Melioidosis | 0.494 | 0.0344 | 67 | Yes |
| Campylobacter coli | Bacteria | Enteritis, Campylobacteriosis | 2.12 | 0.144 | 16 | Yes |
| Campylobacter jejuni | Bacteria | Enteritis | 2.12 | 0.1444 | 16 | Yes |
| Chlamydophila abortus | Bacteria | Chlamydiosis | 0.283 | - | 0 | No |
| Chlamydophila felis | Bacteria | Chlamydiosis | 0.283 | 0.0384 | 60 | No |
| Chlamydophila pneumoniae | Bacteria | Chlamydiosis | 0.283 | 0.039 | 59 | Yes |
| Clostridium botulinum | Bacteria | Botulism | 1.975 | 0.0412 | 56 | Yes |
| Clostridium perfringens | Bacteria | sepsis, toxins, food poisoning | 5 | 0.06 | 38 | Yes |
| Coccidioides immitis | Fungal Spore | Coccidioidomycosis | 3.464 | - | - | Yes |
| Coxiella burnetii | Bacteria | Q Fever | 0.283 | 0.1535 | 15 | Yes |
| Cryptococcus neoformans | Fungi | Cryptococcosis | 4.899 | 0.0167 | 138 | Yes |
| Dermatophilus congolensis | Bacteria | dermatophilosis, mud fever | 1 | - | - | No |
| Epidermophyton | Fungi | Dermatophytosis | 8.5 | - | - | Yes |
| Leptospira spp. | Spirochete | Leptospirosis | 0.1 | NA | - | Yes |
| Listeria monocytogenes | Bacteria | Listeriosis | 0.707 | 0.0127 | 181 | Yes |
| Louping Ill (LIV) | Virus | Encephalomyelitis | 0.05 | 0.003289 | 700 | Yes |
| Microsporum spp. | Fungal Spore | Dermatophytosis | 2.96 | - | - | Yes |
| Mycobacterium avium | Bacteria | Paratuberculosis, Johne's Disease | 1.118 | 0.04387 | 52 | Yes |
| Mycobacterium bovis | Bacteria | Tuberculosis | 0.637 | 0.181 | 13 | Yes |
| Pseudorabies (PRV) | Virus | Aujeszky's Disease Virus (ADV) | 0.194 | 0.0676 | 34 | No |
| Rabies virus | Virus | rabies | 0.07 | 0.219 | 11 | Yes |
| Salmonella enteritidis | Bacteria | Salmonellosis | 0.81 | 0.221 | 10 | Yes |
| Salmonella typhi | Bacteria | Salmonellosis | 0.81 | 0.1467 | 16 | Yes |
| Staphylococcus aureus (MRSA) | Bacteria | MRSA, various infections | 0.866 | 0.596 | 4 | Yes |
| Streptococcus pyogenes | Bacteria | fever | 0.894 | 1.561 | 1 | Yes |
| Trichophyton spp. | Fungal Spore | Dermatophytosis | 7 | 0.00411 | 560 | Yes |

Table 8A: UV24 Removal Rates for Cow Pathogens

| PATHOGEN | Mean | MERV 6 | UV k | Single Pass | Total Removal |
|------------------------------|--------------------|---------------|-----------------------|-----------------|---------------|
| | Dia. μm | Filtration, % | m^2/J | UV Kill Rate, % | Rate, % |
| Actinomyces bovis | 0.5 | 6.89 | - | - | - |
| Bacillus anthracis | 1.118 | 18.79 | 0.0270 | 99.53 | 99.61 |
| Bovine Coronavirus (BCV) | 0.113 | 6.29 | 0.3770 | 100.00 | 100.00 |
| Brucella abortus | 0.57 | 8.05 | 0.0307 | 99.77 | 99.79 |
| Brucella canis | 0.566 | 7.98 | 0.0480 | 99.99 | 99.99 |
| Brucella maris | 0.566 | 7.98 | 0.0470 | 99.99 | 99.99 |
| Brucella melitensis | 0.566 | 7.98 | 0.0470 | 99.99 | 99.99 |
| Brucella neotomae | 0.566 | 7.98 | 0.0470 | 99.99 | 99.99 |
| Brucella ovis | 0.566 | 7.98 | 0.0470 | 99.99 | 99.99 |
| Brucella suis | 0.57 | 8.05 | 0.0470 | 99.99 | 99.99 |
| Burkholderia pseudomallei | 0.494 | 6.80 | 0.0344 | 99.89 | 99.90 |
| Campylobacter coli | 2.12 | 35.03 | 0.1440 | 100.00 | 100.00 |
| Campylobacter jejuni | 2.12 | 35.03 | 0.1444 | 100.00 | 100.00 |
| Chlamydomphila abortus | 0.283 | 4.42 | - | - | - |
| Chlamydomphila felis | 0.283 | 4.42 | 0.0384 | 99.95 | 99.95 |
| Chlamydomphila pneumoniae | 0.283 | 4.42 | 0.0390 | 99.96 | 99.96 |
| Clostridium botulinum | 1.975 | 33.20 | 0.0412 | 99.97 | 99.98 |
| Clostridium perfringens | 5 | 48.83 | 0.0600 | 100.00 | 100.00 |
| Coccidioides immitis | 3.464 | 45.25 | - | - | - |
| Coxiella burnettii | 0.283 | 4.42 | 0.1535 | 100.00 | 100.00 |
| Cryptococcus neoformans | 4.899 | 48.72 | 0.0167 | 96.34 | 98.12 |
| Dermatophilus congolensis | 1 | 16.43 | - | - | - |
| Epidermophyton | 8.5 | 49.96 | - | - | - |
| Leptospira spp. | 0.1 | 6.97 | - | - | - |
| Listeria monocytogenes | 0.707 | 10.57 | 0.0127 | 91.91 | 92.77 |
| Louping Ill (LIV) | 0.05 | 12.45 | 0.0033 | 47.86 | 54.35 |
| Microsporium spp. | 2.96 | 42.60 | - | - | - |
| Mycobacterium avium | 1.118 | 18.79 | 0.0439 | 99.98 | 99.99 |
| Mycobacterium bovis | 0.637 | 9.25 | 0.1810 | 100.00 | 100.00 |
| Pseudorabies (PRV) | 0.194 | 4.47 | 0.0676 | 100.00 | 100.00 |
| Rabies virus | 0.07 | 9.46 | 0.2190 | 100.00 | 100.00 |
| Salmonella enteritidis | 0.81 | 12.59 | 0.2210 | 100.00 | 100.00 |
| Salmonella typhi | 0.81 | 12.59 | 0.1467 | 100.00 | 100.00 |
| Staphylococcus aureus (MRSA) | 0.866 | 13.72 | 0.5960 | 100.00 | 100.00 |
| Streptococcus pyogenes | 0.894 | 14.28 | 1.5610 | 100.00 | 100.00 |
| Trichophyton spp. | 7 | 49.82 | 0.0041 | 55.68 | 77.76 |

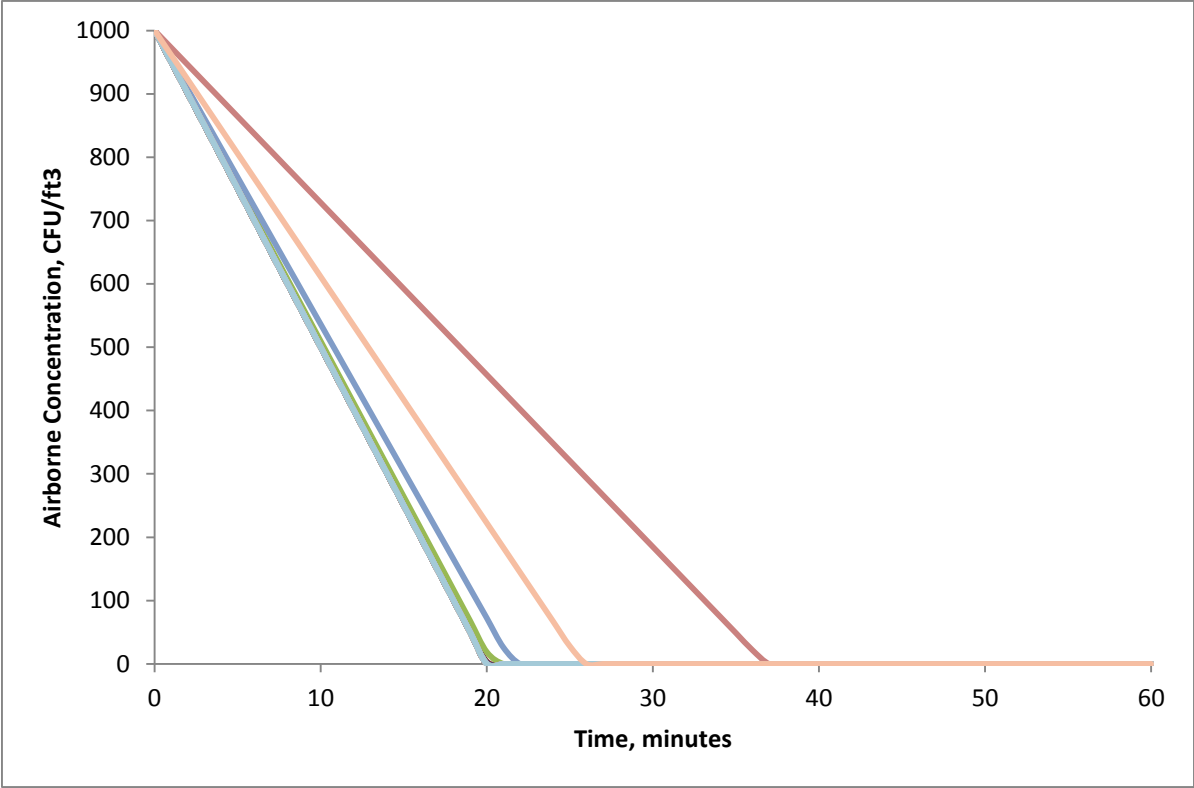


Figure 8: Removal of Bovine pathogens by the UV24 unit in a room of 800 ft³ volume with no outside air. Only microbes from Table 8A with known UV rate constants are included.

Airborne Diseases of Swine

Table 9 lists all of the potentially airborne zoonotic pathogens of swine. Table 9A summarizes the calculated removal rates for all the pathogens in Table 9, except where the UV rate constant is unknown. Figure 9 illustrates the removal rates of the pathogens in Table 9A in a model room of 800 ft³ with no outside air and this graph only includes those pathogens with known UV rate constants. It can be observed in Figure 9 that all pathogens are removed to near-zero levels within about one hour.

Table 9: Potentially Airborne Pathogens and Allergens of Swine

| PATHOGEN or DISEASE | GROUP | DISEASE | Mean Dia. μm | UV k m^2/J | UV D90 J/m^2 | Trans to Human |
|----------------------------------------|--------------|-----------------------------------|-----------------------------------------------|--------------------------------------------------|----------------------------------------------------|---------------------------|
| Arterivirus | V | EAV, PRRD | 0.05 | - | - | No |
| Bovine Coronavirus (BCV) | V | respiratory infections | 0.113 | 0.377 | 6 | No |
| Brucella abortus | B | Brucellosis, undulant fever | 0.57 | 0.0307 | 75 | Yes |
| Brucella melitensis | B | Brucellosis | 0.566 | 0.047 | 49 | Yes |
| Clostridium perfringens | B | sepsis, toxins, food poisoning | 5 | 0.06 | 38 | Yes |
| Mycobacterium avium | B | Paratuberculosis, Johne's Disease | 1.118 | 0.04387 | 52 | Yes |
| Mycoplasma hyopneumoniae | B | chronic infectious pneumoniae | 0.177 | 0.284 | 8 | No |
| Porcine Respiratory Coronavirus (PRCV) | V | respiratory disease | 0.14 | 0.1706 | 13 | No |
| Pseudorabies (PRV) | V | Aujeszky's Disease Virus (ADV) | 0.194 | 0.0676 | 34 | No |
| Salmonella enteritidis | B | Salmonellosis | 0.81 | 0.221 | 10 | Yes |
| Salmonella typhi | B | Salmonellosis | 0.81 | 0.1467 | 16 | Yes |
| Swine Influenza | V | H1N1 flu | 0.1 | 0.098 | 23 | Yes |

Table 9A: UV24 Removal Rates for Swine Pathogens

| PATHOGEN | Mean Dia. μm | MERV 6 Filtration, % | UV k m^2/J | Single Pass UV Kill Rate, % | Total Removal Rate, % |
|----------------------------------------|-----------------------------------------------|---------------------------------|--------------------------------------------------|----------------------------------------|----------------------------------|
| Arterivirus | 0.05 | 12.45 | - | - | - |
| Bovine Coronavirus (BCV) | 0.113 | 6.29 | 0.3770 | 100.00 | 100.00 |
| Brucella abortus | 0.57 | 8.05 | 0.0307 | 99.77 | 99.79 |
| Brucella melitensis | 0.566 | 7.98 | 0.0470 | 99.99 | 99.99 |
| Clostridium perfringens | 5 | 48.83 | 0.0600 | 100.00 | 100.00 |
| Mycobacterium avium | 1.118 | 18.79 | 0.0439 | 99.98 | 99.99 |
| Mycoplasma hyopneumoniae | 0.177 | 4.64 | 0.2840 | 100.00 | 100.00 |
| Porcine Respiratory Coronavirus (PRCV) | 0.14 | 5.34 | 0.1706 | 100.00 | 100.00 |
| Pseudorabies (PRV) | 0.194 | 4.47 | 0.0676 | 100.00 | 100.00 |
| Salmonella enteritidis | 0.81 | 12.59 | 0.2210 | 100.00 | 100.00 |
| Salmonella typhi | 0.81 | 12.59 | 0.1467 | 100.00 | 100.00 |
| Swine Influenza | 0.1 | 6.97 | 0.0980 | 100.00 | 100.00 |

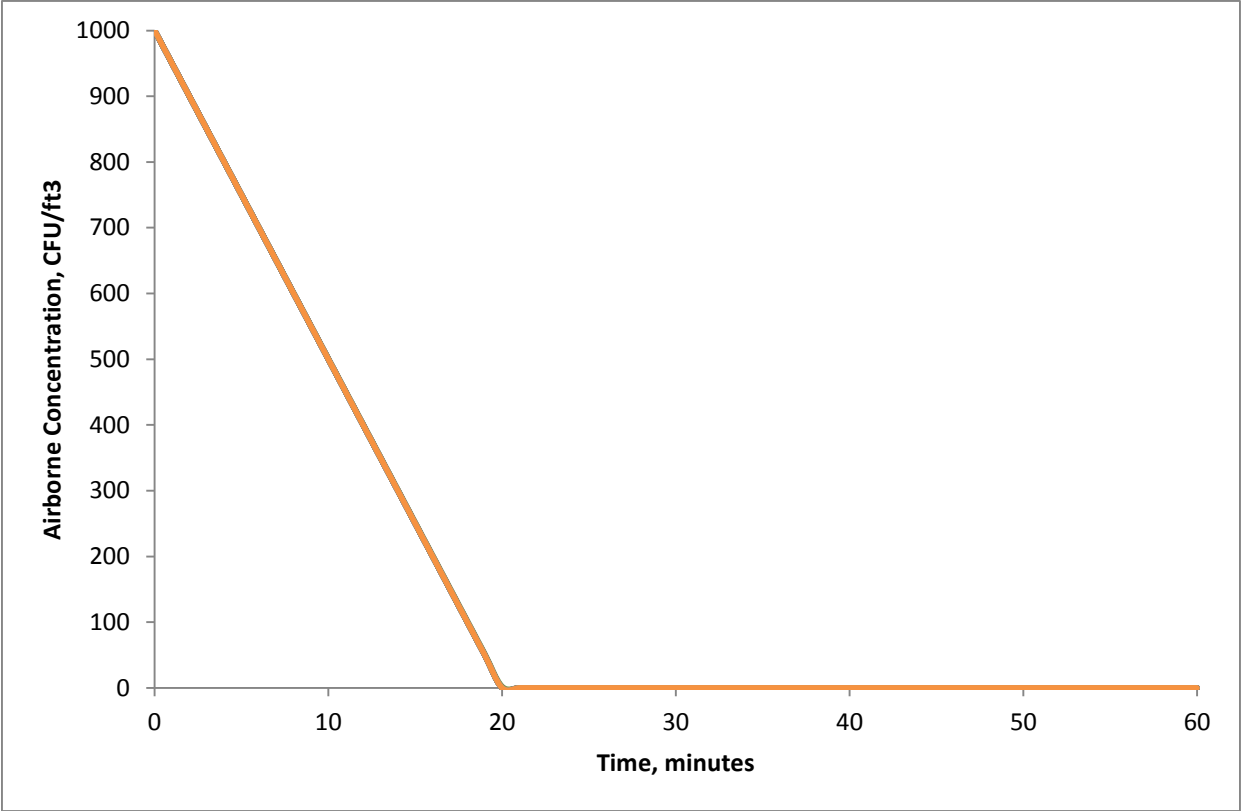


Figure 9: Removal of Swine pathogens by the UV24 unit in a room of 800 ft³ volume with no outside air. Only microbes from Table 9A with known UV rate constants are included.

Airborne Diseases of Reptiles and Amphibians

Table 10 lists all of the potentially airborne zoonotic pathogens of reptiles and amphibians. Table 10A summarizes the calculated removal rates for all the pathogens in Table 10, except where the UV rate constant is unknown. Figure 10 illustrates the removal rates of the pathogens in Table 10A in a model room of 800 ft³ with no outside air and this graph only includes those pathogens with known UV rate constants. It can be observed in Figure 10 that all pathogens are removed to near-zero levels within about one hour.

Table 10: Potentially Airborne Pathogens and Allergens of Reptiles and Amphibians

| PATHOGEN or DISEASE | GROUP | DISEASE | Mean Dia. μm | UV k m^2/J | UV D90 J/m^2 | Trans to Human |
|--------------------------------|--------------|----------------------------------|-----------------------------------------------|--------------------------------------------------|----------------------------------------------------|---------------------------|
| Adenovirus | Virus | hepatitis | 0.093 | 0.026 | 89 | No |
| Batrachochytrium dendrobatidis | Fungi | Chytridiomycosis | 4 | unk | unk | No |
| Calicivirus | Virus | fever, conjunctivitis | 0.034 | 0.0345 | 66.7416 | No |
| Herpesvirus | Virus | Pneumonia, respiratory infection | 0.18 | 0.0568 | 41 | No |
| Mycobacteria | Bacteria | Mycobacteriosis | 0.1 | unk | unk | Yes |
| Mycoplasma agassizii | Bacteria | upper respiratory tract disease | 0.177 | 0.284 | 8 | No |
| Ophidian Paramyxovirus (PMV) | Virus | Pneumonia, respiratory infection | 0.15 | unk | unk | No |
| Reovirus | Virus | Pneumonia, respiratory infection | 0.08 | 0.016 | 143.9116 | Yes |

Table 10A: UV24 Removal Rates for Reptile Pathogens

| PATHOGEN | Mean Dia. μm | MERV 6 Filtration, % | UV k m^2/J | Single Pass UV Kill Rate, % | Total Removal Rate, % |
|--------------------------------|-----------------------------------------------|---------------------------------|--------------------------------------------------|----------------------------------------|----------------------------------|
| Adenovirus | 0.093 | 7.41 | 0.0260 | - | - |
| Batrachochytrium dendrobatidis | 4 | 47.06 | unk | - | - |
| Calicivirus | 0.034 | 16.39 | 0.0345 | 99.89 | 99.91 |
| Herpesvirus | 0.18 | 4.60 | 0.0568 | 100.00 | 100.00 |
| Mycobacteria | 0.1 | 6.97 | unk | - | - |
| Mycoplasma agassizii | 0.177 | 4.64 | 0.2840 | 100.00 | 100.00 |
| Ophidian Paramyxovirus (PMV) | 0.15 | 5.09 | unk | - | - |
| Reovirus | 0.08 | 8.44 | 0.0160 | 95.79 | 96.15 |

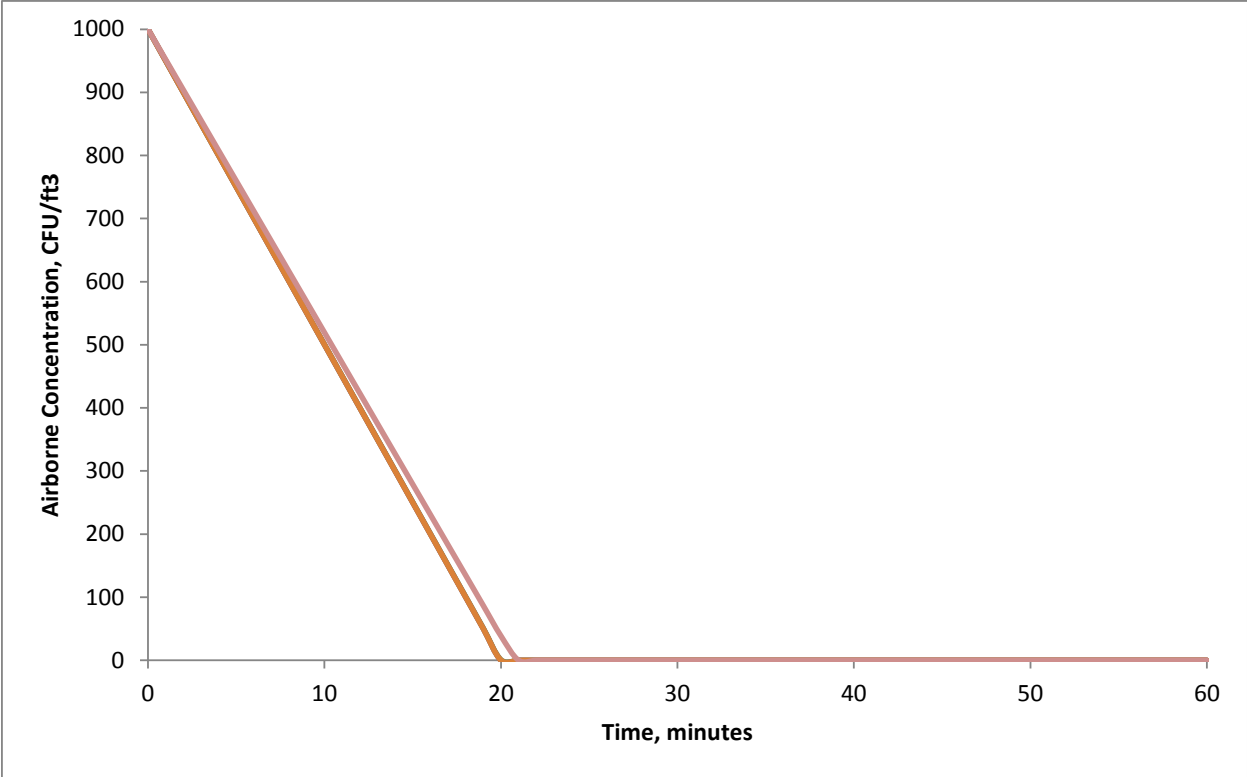


Figure 10: Removal of Reptile and Amphibian pathogens by the UV24 unit in a room of 800 ft³ volume with no outside air. Only microbes from Table 10A with known UV rate constants are included.

References

- AAAAI (2000). "Pollen & Spore Report." , American Academy of Allergy, Asthma and Immunology, Milwaukee.
- AAFA (2005). "*Indoor Air Quality and Allergies.*" Asthma and Allergy Foundation of America, www.aafa.org.
- AAFA (2011). "*Allergy Overview.*" Asthma and Allergy Foundation of America, www.aafa.org.
- AAHA (2006). "AAHA canine vaccine guidelines." , American Animal Hospital Association, Lakewood, CO.
- Abdel-moein, K. A., El-Hariri, M., and Samir, A. (2011). "Methicillin-resistant *Staphylococcus aureus*: An emerging pathogen of pets in Egypt with a public health burden." *Transboundary and Emerging Diseases DOI: 10.1111/j.1865-1682.2011.01273.x*
- Abe, K. (1996). "Effects of reducing relative humidity on fungal viability." *The 7th International Conference on IAQ and Climate*, Nagoya, Japan, 209-214.
- Abraham, G. (1979). "The effect of ultraviolet radiation on the primary transcription of Influenza virus messenger RNAs." *Virology* 97, 177-182.
- Abshire, R. L., and Dunton, H. (1981). "Resistance of selected strains of *Pseudomonas aeruginosa* to low-intensity ultraviolet radiation." *Appl Environ Microbiol* 41(6), 1419-1423.
- ACGIH (2010). "Industrial Ventilation: A Manual of Recommended Practice." , American Conference of Governmental Industrial Hygienists, Cincinnati, OH.
- AIA (2001). *Guidelines for construction and equipment of hospital and medical facilities* Mechanical Standards A. I. o. Architects, ed., Washington
- AIHA (2003). "*ANSI/AIHA Z9.5-2003 Laboratory Ventilation Standard.*" , American Industrial Hygiene Association, Fairfax, VA.
- AKC (2008). "American Kennel Club position statement on proper care." , American Kennel Club, Raleigh, NC.
- AKC (2006). "American Kennel Club position statement on deficiencies in the care and condition of dogs." , American Kennel Club, Raleigh, NC.
- Amman, H. M. (2001). "Is Indoor Mold Contamination a Threat to Health?"
- ANSI (1992). *American national standard for laboratory ventilation.* ANSI/AIHA American National Standards Institute, New York.
- APDT (2003). "Code of professional conduct and responsibility." , Association of Pet Dog Trainers, Greenville, SC.
- APIC (1995). "APIC guideline for handwashing and antisepsis in health care settings." *Am J Inf Control* 23, 251-269.
- ASHRAE (1999). *Handbook of Applications.* ASHRAE, Atlanta.
- ASHRAE (2003). *HVAC Design Manual for Hospitals and Clinics.* American Society of Heating, Ventilating, and Air Conditioning Engineers, Atlanta.
- ASPCA (2009). "State Shelter Regulations." , American Society for the Prevention of Cruelty to Animals, New York, NY.
- ASPCA (2009a). "Shelter Regulations." , American Society for the Prevention of Cruelty to Animals, New York, NY.
- Asthana, A., and Tuveson, R. W. (1992). "Effects of UV and phototoxins on selected fungal pathogens of citrus." *Int J Plant Sci* 153(3), 442-452.
- ASV (2010). "Guidelines for Standards of Care in Animal Shelters." , Association of Shelter Veterinarians
- AVMA (2009). "Backgrounder: Canine Influenza." , American Veterinary Medical Association.
- AVMA (2008a). "AVMA animal welfare principles." , American Veterinary Medical Association.
- AVMA (2008). "AVMA companion animal care guidelines." , American Veterinary Medical Association.
- AZA (2009). "Animal Husbandry and Welfare." , Association of Zoos and Aquariums, Silver Spring, MD.
- AZA (2010). "The Accreditation Standards and Policies." , Association of Zoos and Aquariums, Silver Spring, MD.
- Battigelli, D., Sobsey, M., and Lobe, D. (1993). "The inactivation of hepatitis A virus and other model viruses by UV irradiation." *Wat Sci Technol* 27, 339.
- Beebe, J. M. (1959). "Stability of disseminated aerosols of *Pasteurella tularensis* subjected to simulated solar radiations at various humidities." *Journal of Bacteriology* 78, 18-24.
- Begum, M., Hocking, A., and Miskelly, D. (2009). "Inactivation of food spoilage fungi by ultraviolet (UVC)

- irradiation." *Int J Food Microbiol* 129, 74-77.
- Bell, C., and Kyriakides, A. (1998). *Listeria: A practical approach to the organism and its control in foods*. Blackie Academic & Professional, London.
- Bener, A., Mobayed, H., Sattar, H. A., Al-Mohammed, A., Ibrahim, A. S., and Sabbah, A. (2004). "Pets ownership: Its effect on allergy and respiratory symptoms." *Allerg Immunol* 36, 306-310.
- Bener, A., Galadari, I., and Naser, K. A. (1995). "Pets, allergy and respiratory symptoms in children living in a desert country." *Allergy Immunol* 27, 190-195.
- Benirschke, K., Garner, F. M., and Jones, T. C. (1978). "*Pathology of Laboratory Animals*." Springer-Verlag. New York.
- Benitah, N. (2006). "Canine Nasal Aspergillosis." *Clin Tech Small Anim Pract* 21, 82-88.
- Besch, E. L. (1980). "Environmental quality within animal facilities." *Lab Animal Sci* 30(2), 385-398.
- Bisseru, B. (1967). *Diseases of Man Acquired From His Pets*. J.B. Lippencott Company, Philadelphia.
- Boshoff, H. I. M., Reed, M. B., Barry, C. E., and Mizrahi, V. (2003). "DnaE2 polymerase contributes to in vivo survival and the emergence of drug resistance in *Mycobacterium tuberculosis*." *Cell* 113, 183-193.
- Brickus, L. D., Siquiera, L. F. G., Silveira, M. J., Cardoso, J. N., and Neto, F. R. d. A. (1997). "Characteristics of indoor and outdoor airborne microorganisms in southeastern Brazilian offices." *Healthy Buildings IAQ '97*, 239244.
- Brooke, C. J., and Riley, T. V. (1999). "*Erysipelothrix rhusiopathiae*: Bacteriology, epidemiology and clinical manifestations of an occupational pathogen." *J Med Microbiol* 48, 789-799.
- Burge, H. A. (2002). "An update on pollen and fungal spore aerobiology." *J Allergy Clin Immunol* 110(4), 544-552.
- Butler, R. C., Lund, V., and Carlson, D. A. (1987). "Susceptibility of *Campylobacter jejuni* and *Yersinia enterocolitica* to UV radiation." *Zbl Vet Med B* 29, 129-136.
- CFSPH (2005b). "*Listeriosis*." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2007b). "*Porcine and Ruminant Brucellosis: Brucella suis*." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2007d). "*Bovine tuberculosis*." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2007c). "*Glanders*." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2003). "*Louping Ill*." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2009). "*Influenza*." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2009a). "*Avian Influenza*." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2009d). "*Hendra Virus Infection*." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2007f). "*Anthrax*." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2009b). "*Rabies*." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2010a). "*Dermatophilosis*." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2003a). "*Botulism*." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2009c). "*Swine Influenza*." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2006d). "*Aujeszky's Disease*." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2007a). "*Ovine and Caprine Brucellosis: Brucella melitensis*." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2004). "*Plague*." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.

- CFSPH (2004c). "Coccidioidomycosis." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2007e). "Nipah Virus Infection." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2004b). "Epsilon toxin of *Clostridium perfringens*." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2006c). "Sporotrichosis." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2005). "Zoonotic Chlamydiae from Mammals." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2005a). "Dermatophytosis." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2007). "Canine Brucellosis: *Brucella canis*." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2009e). "Canine Brucellosis: *Brucella canis*." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2005). "Cryptococcosis." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2005c). "Campylobacteriosis." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2006a). "Rat Bite Fever." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2010). "High Pathogenicity Avian Influenza." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2006). "Methicillin Resistant *Staphylococcus aureus*." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2006b). "Lymphocytic Choriomeningitis." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2010b). "Bovine Brucellosis: *Brucella abortus*." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- CFSPH (2004a). "Tularemia." Center for Food Security and Public Health / Iowa State University / <http://www.cfsph.iastate.edu/>.
- Chang, C. P., New, A. E., Taylor, J. F., and Chiang, H. S. (1976). "Influenza virus isolations from dogs during a human epidemic in Taiwan." *Int J Zoon* 3, 61-64.
- Chang, J. C. H., Ossoff, S. F., Lobe, D. C., Dorfman, M. H., Dumais, C. M., Qualls, R. G., and Johnson, J. D. (1985). "UV inactivation of pathogenic and indicator microorganisms." *Appl & Environ Microbiol* 49(6), 1361-1365.
- Chick, E. W., A.B. Hudnell, J., and Sharp, D. G. (1963). "Ultraviolet sensitivity of fungi associated with mycotic keratitis and other mycoses." *Sabouviad* 2(4), 195-200.
- Childs, J. E., Glass, G. E., Korch, G. W., Ksiazek, T. G., and ., J. W. L. (1992). "Lymphocytic choriomeningitis virus infection and house mouse (*Mus musculus*) distribution in urban Baltimore." *Am J Trop Med Hyg* 47(1), 27-34.
- Clark, J. D., Gebhart, G. F., Gonder, J. C., Keeling, M. E., and Kohn, D. F. (1996). "The 1996 Guide for the Care and Use of Laboratory Animals." *ILAR Journal* 38(1), 41-48.
- Collins, F. M. (1971). "Relative susceptibility of acid-fast and non-acid fast bacteria to ultraviolet light." *Appl Microbiol* 21, 411-413.
- Cornelis, J. J., and Rommelaere, J. (1982). "Direct and indirect effects of ultraviolet light on the mutagenesis of parvovirus H-1 in human cells." *EMBO J* 1(6), 693-699.
- Corona-Barrera E, S. D., Murray B, Thomson JR. . (2004). "Efficacy of seven disinfectant sanitisers on field isolates of *Brachyspira pilosicoli*." *Vet Rec* 154(15), 473-474.
- Council, E. (1986). "European convention for the protection of vertebrate animals used for experimental and other scientific purposes." , European Council, Council of Europe, Strasbourg.
- Crane, J. T. (1994). "Biological laboratory ventilation and architectural and mechanical implications of biological safety cabinet selection, location, and venting." *ASHRAE Transactions* 100, 1257-1265.
- CVMA (2004). "Humans training methods for dogs." , Canadian Veterinary Medical Association, Ottawa, ON.

- CVMA (2007). "A code of practice for Canadian kennel operations." , Canadian Veterinary Medical Association, Ottawa, ON.
- Dalton, A. J., and Haguenu, F. (1973). *Ultrastructure of Animal Viruses and Bacteriophages: An Atlas*. Academic Press, New York.
- Damani, N. N. (1997). *Manual of Infection Control Procedures*. Greenwich Medical Media, Ltd., London.
- David, H. L. (1973). "Response of mycobacteria to ultraviolet radiation." *Am Rev Resp Dis* 108, 1175-1184.
- David, H. L., Jones, W. D., and Newman, C. M. (1971). "Ultraviolet light inactivation and photoreactivation in the mycobacteria." *Infect and Immun* 4, 318-319.
- de Roda Husman, A. M., Bijkerk, P., Lodder, W., Berg, H. v. d., Pribil, W., Cabaj, A., Gehringer, P., Sommer, R., and Duizer, E. (2004). "Calicivirus Inactivation by Nonionizing (253.7-Nanometer-Wavelength [UV]) and Ionizing (Gamma) Radiation." *Appl Environ Microbiol* 70(9), 5089-5093.
- Deshmukh, D., and Pomeroy, B. (1968). "Ultraviolet inactivation and photoreactivation of avian viruses." *Sci J Minnesota Agricult Exp Station Paper No. 6744*
- DHHS (1993). "Biosafety in Microbiological and Biomedical Laboratories." , U.S. Department of Health and Human Services, Cincinnati, OH.
- Dobly, A., Cochez, C., Goossens, E., Bosschere, H. D., Hansen, P., Roels, S., and Heyman, P. (2012). "Sero-epidemiological study of the presence of hantaviruses in domestic dogs and cats from Belgium." *Res Vet Sci* 92(1), 221-224.
- Dolman, P. J., and Dobrogowski, M. J. (1989). "Contact lens disinfection by ultraviolet light." *Am J Ophthalmol* 108(6), 665-669.
- Driscoll, C. A., Macdonald, D. W., and O'Brien, S. J. (2009). "From wild animals to domestic pets, an evolutionary view of domestication." *PNAS* 106, 9971-9978.
- Duhamel, G. E., Kinyon, J. M., Mathiesen, M. R., Murphy, D. P., and Walter, D. (1998). "In vitro activity of four antimicrobial agents against North American isolates of porcine *Serpulina pilosicoli*." *J Vet Diagn Invest* 10, 350-356.
- Dvorak, G. (2008). "Disinfection 101." , Center for Food Security and Public Health, Iowa State University.
- Etzel, R. A., Balk, S. J., Bearer, C. F., Miller, M. D., Shannon, M. W., and Shea, K. M. (1998). "Toxic effects of indoor molds." *Pediatrics* 101(4), 712-714.
- Evermann JF, M. A., Smith AW, Skilling DE, Ott RL. (1985). "Isolation and identification of caliciviruses from dogs with enteric infections." *American Journal of Veterinary Research* 46(1), 218-220.
- FASS (1999). "Guide for the care and use of agricultural animals in agricultural research and teaching." , Federation of Animal Science Societies, Champaign, IL.
- FCHS (2010). "4-b-1 Cleaning and Maintenance of Dog Kennels." , Franklin County Humane Society, Frankfort, KY.
- Fiennes, R. (1978). *Zoonoses and the Origins and Ecology of Human Disease*. Academic Press, London.
- Flannigan, B., and Hunter, C. A. (1988). *Factors affecting airborne moulds in domestic dwellings* Indoor Air and Ambient Air Quality R. Perry and P. W. Kirk, eds., 461-468.
- Flannigan, B., McCabe, E. M., Jupe, S. V., and Jeffrey, I. G. (1993). "Mycological and acralogical investigation of complaint and noncomplaint houses in Sctland." *6th International Conference on Indoor Air Quality and Climate*, Helsinki, Finland
- Flannigan, B., McCabe, E. M., and McGarry, F. (1991). *Allergenic and toxigenic micro-organisms in houses* Pathogens in the Environment B. Austin, ed., Blackwell Scientific Publications, Oxford
- Flannigan, B., McEvoy, E. M., and McGarry, F. (1999). "Investigation of airborne and surface bacteria in homes." *Indoor Air 99 : Proceedings of the 8th International Conference on Indoor Air Quality and Climate*, Edinburgh, Scotland, 884-889.
- Flannigan, B., Samson, R. A., and Miller, J. D. (2001). *Microorganisms in home and indoor work environments*. Taylor and Francis. Andover, Hants, UK.
- Fleming, D. O., Richardson, J. H., Tulis, J. J., and Vesley, D. (1995). *Laboratory Safety Principles and Practices, Second Edition*. ASM Press. Washington, D.C.
- Fletcher, L. (2004). "The influence of relative humidity on the UV susceptibility of airborne gram negative bacteria." *IUVA News* 6(1), 12-19.
- Forbes, L. B. (1990). "*Brucella abortus* infection in 14 farm dogs." *J Am Vet Med Assoc* 196(6), 911-916.
- Fossi, M. (2006). "Epidemiological aspects and improved differential diagnostics of porcine *Brachyspira pilosicoli*." , University of Helsinki, Helsinki.
- Fulton, H. R., and Coblenz, W. W. (1929). "The fungicidal action of ultraviolet radiation." *J Agric Res* 38,

- Furness, G. (1977). "Differential responses of single cells and aggregates of Mycoplasmas to ultraviolet irradiation." *Appl Microbiol* 18(3), 360-364.
- Gates, F. L. (1934). "Results of irradiating *Staphylococcus aureus* bacteriophage with monochromatic ultraviolet light." *J Exp Med* 60, 179-188.
- Gates, F. L. (1929). "A study of the bactericidal action of ultraviolet light." *J Gen Physiol* 13, 231-260.
- Green, C. F., Favidson, C. S., Scarpino, P. V., and S. G., Gibbs (2004). "Disinfection of selected *Aspergillus* spp. using ultraviolet irradiation." *Can J Microbiol* 50(3), 221-224.
- Green, R., Simpson, A., Custovic, A., Faragher, B., Chapman, M., and Woodcock, A. (1999). "The effect of air filtration on airborne dog allergen." *Allergy* 54, 484-488.
- Greene, C. (1998). *Infectious Diseases of the Dog and Cat*. W.B. Saunders Company, Philadelphia.
- Gritz, D. C., Lee, T. Y., McDonnell, P. J., Shih, K., and Baron, N. (1990). "Ultraviolet radiation for the sterilization of contact lenses." *CLAO J* 16(4), 294-298.
- Grmek, M. D. (1989). *Diseases in the Ancient Greek World*. The Johns Hopkins University Press, Baltimore, MD.
- Hansen, A. K. (2000). *Handbook of Laboratory Animal Bacteriology*. CRC Press, Boca Raton.
- Harder, T. C., and Vahlenkamp, T. W. (2010). "Influenza virus infections in dogs and cats." *Vet Immunol Immunopathol* 134(1-2), 54-60.
- Harvey, C. E. (1984). "Nasal aspergillosis and penicilliosis in dogs: results of treatment with thiabendazole." *J Am Vet Med Assoc* 184(1), 48-50.
- Hijnen, W. A. M., Beerendonk, E. F., and Medema, G. J. (2006). "Inactivation credit of UV irradiation for viruses, bacteria and protozoan (oo)cysts in water: A review." *Wat Res* 40, 3-22.
- Hinton, M., and Bale, M. J. (1991). *Bacterial pathogens in domesticated animals and their environment* Pathogens in the Environment B. Austin, ed., Blackwell Scientific Publications, Oxford
- Hollaender, A. (1955). *Radiation Biology, Volume II: Ultraviolet and Related Radiations*. McGraw-Hill, New York.
- Hollaender, A., and Oliphant, J. W. (1944). "The inactivating effect of monochromatic ultraviolet radiation on influenza virus." *J Bact* 48(4), 447-454.
- Howard, D. H., and Howard, L. F. (1983). *Fungi pathogenic for Humans and Animals*. Marcel Dekker, Inc., New York.
- HSUS (2010). "General staffing recommendations for kennel caretakers." , Humane Society of the United States, Washington, DC.
- Hull, T. G. (1963). *Diseases Transmitted from Animals to Man*. Charles C. Thomas, Springfield, IL.
- Hyvarinen, A., Reponen, T., Husman, T., Ruuskanen, J., and Nevalainen, A. (1993). "Composition of fungal flora in mold problem houses determined with four different methods." *IAQ '93, Helsinki*, 273-278.
- Inglis, T. J., and Sagripanti, J. L. (2006). "Environmental factors that affect the survival and persistence of *Burkholderia pseudomallei*." *Appl Environ Microbiol* 72, 6865-6875.
- Jaakkola, J. J., Jaakkola, N., Piipari, R., and Jaakkola, M. S. (2002). "Pets, parental atopy and asthma in adults." *J Allergy Clin Immunol* 109, 784-788.
- Jensen, M. M. (1964). "Inactivation of airborne viruses by ultraviolet irradiation." *Applied Microbiology* 12(5), 418-420.
- Jepson, J. D. (1973). "Disinfection of water supplies by ultraviolet radiation." *Wat Treat Exam* 22, 175-193.
- Johanning, E., Landsbergis, P., Gareis, M., Yang, C. S., and Olmsted, E. (1999). "Clinical Experience and Results of a Sentinel Health Investigation Related to Indoor Fungal Exposure." *Environ Health Perspect* 107(Suppl. 3), 489-494.
- Kapur, V., Whittam, T. S., and Musser, J. M. (1994). "Is *Mycobacterium tuberculosis* 15,000 years old?" *J Infect Dis* 170, 1348-1349.
- Kemp, P. C., Neumeister, H. G., Kircheis, U., Schleibinger, H., Franklin, P., and Ruden, H. (1997). "Fungal genera in an office building with a central HVAC system in an Australian mediterranean climate." *Healthy Buildings/IAQ '97*, Bethesda, MD, 257-260.
- Kemp, P. C., Neumeister-Kemp, H. G., Nickelmann, A., and Murray, F. (1999). "Fungi in the dust extracted from fabric covered furnishings: Preliminary results during method standardisation." *Indoor Air 99 : Proceedings of the 8th International Conference on Indoor Air Quality and Climate*, Edinburgh, Scotland, 890-891.

- Kim, T., Silva, J. L., and Chen, T. C. (2002). "Effects of UV irradiation on selected pathogens in peptone water and on stainless steel and chicken meat." *J Food Prot* 65(7), 1142-1145.
- Klanova, K., and Drahonovska, H. (1999). "The concentrations of mixed populations of fungi in indoor air: Rooms with and without mould problems; rooms with and without health complaints." *Indoor Air 99 : Proceedings of the 8th International Conference on Indoor Air Quality and Climate*, Edinburgh, Scotland, 920-924.
- Klein, B., Filon, A. R., vanZeeland, A. A., and vanderEb, A. J. (1994). "Survival of UV-irradiated vaccinia virus in normal and xeroderma pigmentosum fibroblasts; evidence for repair of UV-damaged viral DNA." *Mutat Res* 307(1), 25-32.
- Knudson, G. B. (1986). "Photoreactivation of ultraviolet-irradiated, plasmid-bearing, and plasmid-free strains of *Bacillus anthracis*." *Appl & Environ Microbiol* 52(3), 444-449.
- Ko, G., First, M. W., and Burge, H. A. (2000). "Influence of relative humidity on particle size and UV sensitivity of *Serratia marcescens* and *Mycobacterium bovis* BCG aerosols." *Tuber Lung Dis* 80(4/5), 217-228.
- Kowalski, W., Bahnfleth, W., and Hernandez, M. (2009). "A Genomic Model for Predicting the Ultraviolet Susceptibility of Bacteria and Viruses." *IUVA*, Amsterdam
- Kowalski, W., Bahnfleth, W., and Hernandez, M. (2009). "A Genomic Model for Predicting the Ultraviolet Susceptibility of Viruses." *IUVA News* 11(2), 15-28.
- Kowalski, W., Bahnfleth, W., and Hernandez, M. (2009). "A Genomic Model for the Prediction of Ultraviolet Inactivation Rate Constants for RNA and DNA Viruses." *IUVA*, Boston, MA
- Kowalski, W., and Bahnfleth, W. P. (1998). "Airborne respiratory diseases and technologies for control of microbes." *HPAC* 70(6), 34-48.
- Kowalski, W. J. (2006). *Aerobiological Engineering Handbook: A Guide to Airborne Disease Control Technologies*. McGraw-Hill, New York.
- Kowalski, W. J. (2000). "Indoor mold growth: Health hazards and remediation." *HPAC Engineering* 72(9), 80-83.
- Kowalski, W. J. (2003). *Immune Building Systems Technology*. McGraw-Hill, New York.
- Kowalski, W. J. (2012). *Hospital Airborne Infection Control*. CRC Press/Taylor & Francis, New York.
- Kowalski, W. J., W. P. Bahnfleth, T. S. Whittam (1999). "Filtration of Airborne Microorganisms: Modeling and prediction." *ASHRAE Transactions* 105(2), 4-17. <http://www.engr.psu.edu/ae/wjk/fom.html>.
- Kowalski, W. J. (2009). *Ultraviolet Germicidal Irradiation Handbook: UVGI for Air and Surface Disinfection*. Springer, New York.
- Kowalski, W. J., and Bahnfleth, W. P. (2002a). "Airborne-Microbe Filtration in Indoor Environments." *HPAC Engineering* 74(1), 57-69. <http://www.bio.psu.edu/people/faculty/whittam/research/amf.pdf>.
- Kowalski, W. J., Bahnfleth, W. P., and Carey, D. D. (2002). "Engineering control of airborne disease transmission in animal research laboratories." *Contemporary Topics in Lab Animal Sci* 41(3), 9-17.
- Kowalski, W. J., and Burnett, E. (2001). "Mold and Buildings." *Builder Brief BB0301*, The Pennsylvania Housing Research Center, University Park, PA. <http://www.bio.psu.edu/people/faculty/whittam/research/B0301.pdf>.
- Kurata, H., and Ueno, Y. (1984). *Toxigenic Fungi -- Their Toxins and Health Hazard*. Elsevier, Amsterdam.
- Kutlin, A., Roblin, P. M., Kumar, S., Kohlhoff, S., Bodetti, T., Timms, P., and Hammerschlag, M. R. (2007). "Molecular characterization of *Chlamydomyces pneumoniae* isolates from Western barred bandicoots." *J Med Microbiol* 56(3), 407-417.
- Lacey, J., and Crook, B. (1988). "Fungal and actinomycete spores as pollutants of the workplace and occupational illness." 32, 515-533.
- Lewis, W. H., Vinay, P., and Zenger, V. E. (1983). *Airborne and Allergenic Pollen of North America*. The Johns Hopkins University Press, Baltimore.
- Li, D.-W., and Kendrick, B. (1995). "A year-round comparison of fungal spores in indoor and outdoor air." *Mycologia* 87(2), 190-195.
- Lidwell, O. M., and Lowbury, E. J. (1950). "The survival of bacteria in dust." *Annual Review of Microbiology* 14, 38-43.
- Linton, A. H. (1982). *Microbes, man, and animals: The natural history of microbial interactions*. Wiley & Sons, New York.
- Little, J. S., Kishimoto, R. A., and Canonico, P. G. (1980). "In vitro studies of interaction of rickettsia and macrophages: Effect of ultraviolet light on *Coxiella burnetii* inactivation and macrophage

- enzymes." *Infect Immun* 27(3), 837-841.
- Lucio-Forster, A., Bowman, D. D., Lucio-Martinez, B., Labare, M. P., and Butkus, M. A. (2006). "Inactivation of the Avian Influenza virus (H5N2) in typical domestic wastewater and drinking water treatment systems." *Environ Eng Sci* 23(6), 897-903.
- Luckiesh, M. (1946). *Applications of Germicidal, Erythral and Infrared Energy*. D. Van Nostrand Co., New York.
- Luckiesh, M., Taylor, A. H., Knowles, T., and Leppelmeier, E. T. (1949). "Inactivation of molds by germicidal ultraviolet energy." *Journal of the Franklin Institute* 248(4), 311-325.
- Manian, F. A. (2003). "Asymptomatic nasal carriage of mupirocin-resistant, methicillin-resistant *Staphylococcus aureus* (MRSA) in a pet dog associated with MRSA infection in household contacts." *Clin Infect Dis* 36(2), e36-e28.
- Marks, P. J., Vipond, I. B., Carlisle, D., Deakin, D., Fey, R. E., and Caul, E. O. (2000). "Evidence for airborne transmission of Norwalk-like virus (NLV) in a hotel restaurant." *Epidemiol Infect* 124(3), 481-487.
- Martella, V., and E. Lorusso, N. D., G. Elia, A. Radogna, M. D'Abramo, C. Desario, A. Cavalli, M. Corrente, M. Camero, C. A. Germinario, K. Bányai, B. Di Martino, F. Marsilio, L. E. Carmichael, C. Buonavoglia (2008). "Detection and Molecular Characterization of a Canine Norovirus." *Emerg Inf Dis* 14(8), 1306-1308.
- McCarthy, C., and Schaefer, J. (1974). "Response of *Mycobacterium avium* to ultraviolet irradiation." *Appl Environ Microbiol* 28(1), 151-153.
- McKeown, T. (1988). *The Origins of Human Disease*. Basil Blackwell, Oxford, UK.
- Mesquita, J. R., and Nascimento, M. S. J. (2011). "Gastroenteritis outbreak associated with faecal shedding of Canine Norovirus in a Portugese kennel following introduction of imported dogs from Russia." *Transboundary and Emerging Diseases*(DOI: 10.1111/j.1865-1682.2011.01284.x)
- Mesquita, J. R., Barclay, L., Nascimento, M. S. J., and Vinje, J. (2010). "Novel Norovirus in dogs with diarrhea." *Emerg Inf Dis* 16(6), 980-982.
- Middleton, E., Reed, C. E., and Ellis, E. F. (1983). *Allergy: Principles and Practice, Volume Two*. The C.V. Mosby Company, St. Louis.
- Miller, J. D., Laflamme, A. M., Sobol, Y., Lafontaine, P., and Greenlaugh, R. (1988). "Fungi and Fungal Products in some Canadian Houses." *Intl Biodeterioration* 24, 103-120.
- Miller, L., and Hurley, K. (2009). *Infectious Disease Management in Animal Shelters*. Blackwell Publishing, Ames, IA.
- Miller, L., and Zawistowski, S. (2004). *Shelter Medicine for Veterinarians and Staff*. Blackwell Publishing, Ames, IA.
- Mitscherlich, E., and Marth, E. H. (1984). *Microbial Survival in the Environment*. Springer-Verlag, Berlin.
- Mrozek, M., Zillman, U., Nicklas, W., Kraft, V., Meyer, B., Sickel, E., Lehr, B., and Wetzels, A. (1994). "Efficiency of air filter sets for the prevention of airborne infections in laboratory animal houses." *Laboratory Animals* 28, 347-354.
- NACA (2009a). "Training Manual." , National Animal Care and Control Association, Jamaica Plain, MA.
- NACA (2009). "Determining kennel staffing needs." , National Animal Care and Control Association, Jamaica Plain, MA.
- Nagy, R. (1964). "Application and measurement of ultraviolet radiation." *AIHA J* 25, 274-281.
- Nakamura, H. (1987). "Sterilization efficacy of ultraviolet irradiation on microbial aerosols under dynamic airflow by experimental air conditioning systems." *Bull Tokyo Med Dent Univ* 34(2), 25-40.
- NASPHV (2008). "Zoonotic disease prevention in veterinary personnel." , National Animal Care and Control Association, Jamaica Plain, MA.
- NASPHV (2011). "Compendium of Measures to Prevent Disease Associated with Animals in Public Settings." , National Association of State Public Health Veterinarians, Inc., Nashville, TN.
- NCBI (2009). "Entrez Genome." National Center for Biotechnology Information.
- Nevalainen, A., Reponen, T., Heinonen-Tanski, H., and Raunemaa, T. (1991). *Indoor air bacteria in apartment homes before and after occupancy* IAQ '91 , Healthy Buildings/IAQ '91, Washington
- Nicholson, W., and Galeano, B. (2003). "UV Resistance of *Bacillus anthracis* Spores Revisited: Validation of *Bacillus subtilis* Spores as UV Surrogates for Spores of *B. anthracis* Sterne." *Appl Environ Microbiol* 69(2), 1327-1330.
- Nicklas, W., Homberger, F. R., Illgen-Wilcke, B., Jacobi, K., Kraft, V., Kunstyr, I., Mahler, M., Meyer, H., and Pohlmeier-Esch, G. (1999). "Implications of infectious agents on results of laboratory animal

- experiments." *Laboratory Animals* 33(Suppl.1), S1:39-S1:87.
- Nielsen, K. F., Gravesen, S., Nielsen, P. A., Andersen, B., Thrane, U., and Frisvad, J. C. (1999). "Production of mycotoxins on artificially and naturally infested building materials." *Mycopathologia* 145, 43-56.
- Nowotny, N., Deutz, A., Fuchs, K., Schuller, W., Hinterdorfer, F., Auer, H., and Aspöck, H. (1997). "Prevalence of swine influenza and other viral, bacterial, and parasitic zoonoses in veterinarians." *J Inf Dis* 176, 1414-1415.
- NRC (1996). "Guide for the Care and Use of Laboratory Animals." , N. R. Council, ed. National Academy Press. Washington, D.C.
- Ntafis, V., Xylouri, E., Radogna, A., Buonavoglia, C., and Martella, V. (2010). "Outbreak of Canine Norovirus Infection in Young Dogs." *J Clin Microbiol* 48(7), 2605-2608.
- Nuanualsuwan, S., Mariam, T., Himathongkham, S., and Cliver, D. O. (2002). "Ultraviolet inactivation of Feline Calicivirus, Human Enteric Viruses, and coliphages." *Photochem Photobiol* 76(4), 406-410.
- Nwachuku, N., Gerba, C. P., Oswald, A., and Mashadi, F. D. (2005). "Comparative Inactivation of Adenovirus Serotypes by UV Light Disinfection." *Appl Environ Microbiol* 71(9), 5633-5636.
- Ortega, H. G., Daroowalla, F., Peterson, E. L., Lewis, D., Berardinelli, S., Jones, W., Kreiss, K., and Weissman, D. N. (2001). "Respiratory symptoms among crab processing workers in Alaska: epidemiological and environmental assessment." *Am J Ind Med* 39(6), 598-607.
- OSU (2007). "OSU Small Animal Hospital Housekeeping and Cleaning Procedures." , Ohio State University, Corvallis, OR.
- OSU (2009). "Is airborne fungus a danger to pets or animals?" , Oregon State University, www.kval.com/news/40253267.html
- Peccia, J., and Hernandez, M. (2002). "UV-induced inactivation rates for airborne *Mycobacterium bovis* BCG." *J Occup Environ Hyg* 1(7), 430-435.
- Petersen, C. A., Dvorak, G., and STeneroden, K. (2008). *Maddie's Infection Control Manual for Animal Shelters*. Iowa State University, Center for Food Security and Public Health, Ames, IA.
- Pillai, S. D., and Ricke, S. C. (2002). "Bioaerosols from municipal and animal wastes: background and contemporary issues." *Can J Microbiol* 48(8), 681-696.
- Pratelli, A. (2006). "Genetic evolution of canine coronavirus and recent advances in prophylaxis." *Vet Res* 37(2), 191-200.
- Raoult, D., and Drancourt, M. (2008). *Paleomicrobiology: Past Human Infections*. Springer,
- Ren, J., Guo, R. R., and Wang, X. (1993). "Indoor airborne bacteria concentrations and respiratory disease in old and new rural dwelling." *Proceedings of the 6th International Conference on Indoor Air Quality and Climate*, Helsinki, Finland, 207-212.
- Reponen, T., Hyvarinen, A., Ruuskanen, J., Raunemaa, T., and Nevalainen, A. (1993). "Size distribution of fungal spores in houses with mold problems." *Proceedings of the 6th International Conference on Indoor Air Quality and Climate*, Helsinki, Finland, 243-248.
- Richmond, J. Y., and McKinney, R. W. (1999). "Biosafety in Microbiological and Biomedical Laboratories, 4th Edition,." US Government Printing Office. Washington, D.C.
- Riley, R. L., Knight, M., and Middlebrook, G. (1976). "Ultraviolet susceptibility of BCG and virulent tubercle bacilli." *Am Rev Resp Dis* 113, 413-418.
- Ross, L. J. N., Wildy, P., and Cameron, K. R. (1971). "Formation of small plaques by Herpes viruses irradiated with ultraviolet light." *Virology* 45, 808-812.
- Ruys, T. (1990). *Handbook of Facilities Planning Volume I: Laboratory Facilities*. Van Nostrand Reinhold, New York.
- Ryan, K. J. (1994). *Sherris Medical Microbiology*. Appleton & Lange, Norwalk.
- Rylander, R. (2007). "Endotoxin in the air: Good or bad for you?" *Clin Pulm Med* 14, 140-147.
- Rylander, R., and Etzel, E. (1999). "Introduction and Summary: Workshop on Children's Health and Indoor Mold Exposure." *Environ Health Perspect* 107(Suppl. 3), 465-472.
- Scott, R., and Yang, C. (1997). "Comparison of successful and unsuccessful *Stachybotrys chartarum* remediation projects." *Healthy Buildings/IAQ '97*, Bethesda, MD, 269.
- Sharp, G. (1939). "The lethal action of short ultraviolet rays on several common pathogenic bacteria." *J Bact* 37, 447-459.
- Sharp, G. (1940). "The effects of ultraviolet light on bacteria suspended in air." *J Bact* 38, 535-547.
- Shin, G. (2008). "Inactivation of *Mycobacterium avium* complex by UV irradiation." *Appl Environ Microbiol* 74(22), 7067-7069.

- Skernivitz, S. (2012). "Dogs may harbor, transmit human norovirus." *DVM* 43(3), 8.
- Smith, J. E., and Moss, M. O. (1985). *Mycotoxins: Formation, Analysis and Significance*. John Wiley and Sons, Chichester.
- Smith, J. M. B. (1989). *Opportunistic mycoses of man and other animals*. BPC Wheatons, Exeter.
- Sorensen, K. N., Clemons, K. V., and Stevens, D. A. (1999). "Murine models of blastomycosis, coccidioidomycosis, and histoplasmosis." *Mycopathologia* 146, 53-65.
- Spindel, M., and Makolinski, K. (2008). "Shelter Medicine Resources & Protocols." , American Society for the Prevention of Cruelty to Animals
- Stevenson, G. W. (1999). "*Brachyspira (Serpulina) pilosicoli* and intestinal spirochetosis: How much do we know?" *Swine Health Prod* 7(6), 287-291.
- Sturm, E., Gates, F. L., and Murphy, J. B. (1932). "Properties of the causative agent of a chicken tumor. II. The inactivation of the tumor-producing agent by monochromatic ultra-violet light." *J Exp Med* 55, 441-444.
- Sutton, D. A., Fothergill, A. W., and Rinaldi, M. G. (1998). *Guide to Clinically Significant Fungi*. Williams & Wilkins, Baltimore.
- Suzuki, K., Kayaba, K., Tanuma, T., Kitazawa, J., and Yanagawa, H. (2005). "Respiratory symptoms and hamsters or other pets: a large-sized population survey in Saitama Prefecture." *J Epidemiol* 15, 9-14.
- Thurston-Enriquez, J. A., Haas, C. N., Jacangelo, J., Riley, K., and Gerba, C. P. (2003). "Inactivation of Feline calicivirus and Adenovirus Type 40 by UV radiation." *Appl Environ Microbiol* 69(1), 577-582.
- Tree, J., Adams, M., and Lees, D. (2005). "Disinfection of Feline calicivirus (a surrogate for Norovirus) in wastewaters." *J Appl Microbiol* 98, 155-162.
- Tuffery, A. A. (1995). *Laboratory Animals: An Introduction for Experimenters*. John Wiley & Sons. Chichester.
- UC-Davis (2010). "Facility Design and Animal Housing." , University of California Davis, Sacramento, CA.
- UC-Davis (2009). "Koret Shelter Medicine Program." , University of California Davis, Sacramento, CA.
- USDA (2003). "Caliciviruses of Animals." , United States Department of Agriculture, Fort Collins, CO.
- Van der Hoeden, J. (1964). *Zoonoses*. Elsevier Publishing Company, Amsterdam.
- van Duijkeren, E., Wolfhagen, M. J., Heck, M. E., and Wannet, W. J. (2005). "Transmission of a Pantone-Valentine leucocidin-positive, methicillin-resistant *Staphylococcus aureus* strain between humans and a dog." *J Clin Microbiol* 43(12), 6209-6211.
- VanOsdell, D., and Foarde, K. (2002). "Defining the Effectiveness of UV Lamps Installed in Circulating Air Ductwork." *ARTI-21CR/610-40030-01*, Air-Conditioning and Refrigeration Technology Institute, Arlington, VA.
- Voorhorst, R. (1977). "The human dander atopy. I. The prototype of auto-autopy." *Ann Allergy* 39(3), 205-212.
- Walker, C. M., and Ko, G. (2007). "Effect of ultraviolet germicidal irradiation on viral aerosols." *Environ Sci Technol* 41(15), 5460-5465.
- Wang, Y., and Casadevall, A. (1994). "Decreased susceptibility of melanized *Cryptococcus neoformans* to UV light." *Appl Microb* 60(10), 3864-3866.
- Webb, S. J. (1965). *Bound Water in Biological Integrity*. Charles C. Thomas, Springfield, IL.
- Wedum, A. G. (1961). "Control of laboratory airborne infection." *Bacter Rev* 25, 210-216.
- Weiss, M., and Horzinek, M. C. (1986). "Resistance of Berne virus to physical and chemical treatment." *Vet Microbiol* 11, 41-49.
- WHO (2009). "WHO Guidelines on Hand Hygiene in Health Care: A Summary." , World Health Organization, Geneva.
- Wick, J. Y. (2012). "Global Warming: Climate Change Impacts Allergy Season." *Pharmacy Times* April(26,28)
- Wilson, B., Roessler, P., vanDellen, E., Abbaszadegan, M., and Gerba, C. (1992). "Coliphage MS-2 as a UV water disinfection efficacy test surrogate for bacterial and viral pathogens." *Proceedings of the AWWA Wat Qual Technol Conf*, Denver, CO
- Wright, D. N., G.D.Bailey, and M.T.Hatch (1968). "Survival of airborne *Mycoplasma* as affected by relative humidity." *Journal of Bacteriology* 95(1), 251-252.
- Zahl, P. A., Koller, L. R., and Haskins, C. P. (1939). "The effects of ultraviolet radiation on spores of the fungus *Aspergillus niger*." *J Gen Physiol* 16, 221-235.

- Zavadova, Z. (1971). "Host-cell repair of vaccinia virus and of double stranded RNA of encephalomyocarditis virus." *Nature (London) New Biol* 233, 123.
- Zealand, N. (1998). "Code of recommendations and minimum standards for the welfare of dogs." , New Zealand Ministry of Agriculture, Animal Welfare Advisory Committee, Wellington, New Zealand.

Appendix A: Nonairborne Zoonotic Pathogens

Appendix A: Nonairborne Zoonotic Pathogens and Allergens

| PATHOGEN | Group | Host | DISEASE |
|--------------------------------------------|--------------|-------------|--------------------------------|
| African Animal Trypanosomiasis (AAT) | Protozoa | DCR | Nagana, Tsetse Disease |
| African Horse Sickness Virus (AHS) | Virus | DE | African Horse Sickness |
| African Swine Fever virus | Virus | S | ASF |
| Ainivirus (Bunyaviridae Orthobunyavirus) | Virus | O | Aino Virus Infection, Akabane |
| Air Sac Mite | Insect | B | lung and airway disorder |
| Anaplasma phagocytophilum (Ehrlichia equi) | Bacteria | DCREO | Ehrlichiosis, anaplasmosis |
| Ancylostoma spp. | Nematode | DC | Larva Migrans |
| Avian Enteric Reovirus | Virus | P | Malabsorption syndrome |
| Avian Pathogenic E. coli (APEC) | Bacteria | P | intestinal diseases |
| Avian Reovirus | Virus | P | viral arthritis |
| Avibacterium paragallinarum | Bacteria | P | coryza |
| Babesia gibsoni | Protozoa | DC | hemolytic anemia |
| Bartonella bovis | Bacteria | E | endocarditis |
| Bartonella clarridgeiae | Bacteria | C | Cat Scratch Fever |
| Bartonella henselae | Bacteria | C | Cat Scratch Fever |
| Baylisascaris procyonis | Helminth | D | Verminous myelitis |
| Besnoitia besnoiti | Protozoa | O | Bovine besnoitiosis |
| Birnavirus (Infectious Bursal Disease) | Virus | P | Gumboro disease, IB, IBDV |
| Borna Disease Virus (BDV) | Virus | E | Borna disease |
| Borrelia afzelii | Bacteria | DC | dermatitis, arthritis |
| Borrelia burgdorferi | Bacteria | DCREO | Lyme Disease |
| Borrelia garinii | Bacteria | DC | Meningopolyneuritis |
| Borrelia japonica | Bacteria | DC | Lyme disease |
| Bovine Ephemeral Fever Virus (BEFV) | Virus | O | Ephemeral fever |
| Bovine Papilloma Virus | Virus | E | warts |
| Bovine spongiform encephalopathy (BSE) | Protozoa | O | neurodegenerative disease |
| Canine Herpesvirus (CHV) | Virus | D | Herpes sores, kennel cough |
| Capnocytophaga canimorsus | Bacteria | DC | fulminant sepsis |
| Capripoxvirus | Virus | O | Lumpy Skin Disease |
| Chicken Anemia Virus (CAV) | Virus | P | anemia |
| Crimean-Congo Hemorrhagic Fever (CCHF) | Virus | DRO | Hemorrhagic fever |
| Cryptosporidium canis | Protozoa | D | Cryptosporidiosis |
| Cryptosporidium felis | Protozoa | C | Cryptosporidiosis |
| Cryptosporidium parvum | Protozoa | DCEO | Cryptosporidiosis |
| Cryptosporidium spp. | Protozoa | BS | Cryptosporidiosis |
| Duvenhage virus | Virus | D | bat rabies, human rabies |
| Eastern Equine Encephalomyelitis (EEE) | Virus | DBRE | Encephalitis |
| Echinococcus spp. | Helminth | DR | Echinococcosis |
| Ehrlichia canis | Bacteria | DO | Ehrlichiosis, anaplasmosis |
| Ehrlichia chaffeensis | Bacteria | DRO | Ehrlichiosis, anaplasmosis |
| Ehrlichia ewingii | Bacteria | O | Ehrlichiosis, anaplasmosis |
| Ehrlichia ruminantium | Bacteria | O | Heartwater, cowdriosis |
| Eimeria spp. | Protozoa | P | Coccidiosis |
| Encephalitozoon cuniculi | Protozoa | R | microsporidiosis |
| Encephalitozoon intestinalis | Protozoa | R | microsporidiosis |
| Encephalitozoon hellem | Protozoa | R | microsporidiosis |
| Equine Herpes Virus (EHV) Type 1 | Virus | E | colds, fever, rhinopneumonitis |
| Equine Herpes Virus (EHV) Type 3 | Virus | E | colds, fever, rhinopneumonitis |
| Equine Herpes Virus (EHV) Type 4 | Virus | E | colds, fever, rhinopneumonitis |
| Equine Infectious Anemia Virus | Virus | E | Anemia |

Appendix A: Nonairborne Zoonotic Pathogens and Allergens

| PATHOGEN | Group | Host | DISEASE |
|----------------------------------------|------------|---------|-------------------------------|
| Equine Rabies | Virus | E | rabies |
| Erysipelothrix rhusiopathiae | Bacteria | S | Erysipelas |
| Escherichia coli | Bacteria | DCBPES | various, colibacillosis |
| Escherichia coli F4 | Bacteria | S | diarrhea |
| Escherichia coli F5 | Bacteria | S | diarrhea |
| Feline Immunodeficiency Virus (FIV) | Virus | C | Immunodeficiency |
| Feline Leukemia Virus (FeLV) | Virus | C | Leukemia |
| Feline Spongiform Encephalopathy (FSE) | Protozoa | C | Scrapie, neurodegeneration |
| FMD virus (Aphthovirus Picornaviridae) | Virus | SO | Foot and Mouth Disease |
| Giardia duodenalis | Protozoa | O | Giardiasis |
| Giardia intestinalis | Protozoa | O | Giardiasis |
| Giardia lamblia | Protozoa | O | Giardiasis |
| Giardia spp. | Protozoa | DCBR | Giardiasis |
| Helicobacter pylori | Bacteria | DC | digestive illness |
| Hepatitis E Virus (HEV) | Virus | S | fever |
| Hippobosca longipennis | Insect | DC | Infestation |
| Ibadan shrew virus | Virus | D | rabies-like infection |
| Iso spor a spp. | Protozoa | DC | Coccidiosis |
| Ixodes ricinus | Insect | DCR | Infestation |
| Japanese Encephalitis | Virus | DCRE | Encephalitis |
| Lawsonia intracellularis | Bacteria | S | proliferative enteritis (PE) |
| Leishmania spp. | Protozoa | DCR | Leishmaniasis |
| Leptospira spp. | Spirochete | DCRO | Leptospirosis |
| Lymphoid Leukosis virus | Virus | P | Leukosis |
| Mokola virus | Virus | D | rabies-like infection |
| Mycobacterium lepraemurium | Bacteria | R | leprosy |
| Mycobacterium microti | Bacteria | R | leprosy |
| Mycoplasma haemocanis | Bacteria | DC | haemobartonellosis |
| Neorickettsia helminthoeca | Bacteria | D | Ehrlichiosis |
| Neorickettsia risticii | Bacteria | E | Potomac Horse Fever |
| Neorickettsia sennetsu | Bacteria | O | Ehrlichiosis, anaplasmosis |
| Neospora caninum | Protozoa | D | ascending paralysis |
| Notoedres cati | Insect | R | infestation |
| Ornithonyssus sylviarum | Insect | R | infestation |
| Ornithonyssus bacoti | Insect | R | infestation |
| Pacheco's Disease virus (Herpesvirus) | Virus | B | viral hepatitis |
| Parasitic Feather Mites (Red Mite) | Insect | B | skin infestation |
| Pasteurella multocida (& other spp.) | Bacteria | DCBPRSO | Pasteurellosis, fowl cholera |
| Porcine Circovirus Type 2 | Virus | S | PMWS |
| Porcine Epidemic Diarrhea (PED) virus | Virus | S | Diarrhea, (Coronavirus) |
| Porcine Parvovirus (PPV) | Virus | S | Parvovirus |
| Rickettsia rickettsii | Bacteria | DR | Rocky Mountain Spotted Fever |
| Rift Valley Fever | Virus | DCRO | Infectious Enzootic Hepatitis |
| Rotavirus | Virus | S | diarrhea |
| Salmonella arizona | Bacteria | B | Arizonosis |
| Salmonella bongori | Bacteria | DCRS | Salmonellosis |
| Salmonella enterica | Bacteria | DCRSO | Salmonellosis |
| Salmonella pullorum | Bacteria | P | Pullorum disease |
| Sarcocystis falcata | Protozoa | DCBR | Sarcocystosis |

Appendix A: Nonairborne Zoonotic Pathogens and Allergens

| PATHOGEN | Group | Host | DISEASE |
|-------------------------------------------|----------|-------|-----------------------------------|
| Sarcoptes spp. | Insect | D | Acariasis, scabies |
| Scaly Face (Leg Mite) | Insect | B | Infestation |
| Screwworm Myiasis | Insect | DCR | infestation |
| Streptococcus equi | Bacteria | EO | Strangles, Streptococcosis |
| Strongyloides spp. | Helminth | DC | Larva Migrans |
| Surra | Protozoa | DCRO | Trypanosomosis |
| Swine Vesicular Disease Virus (SVDV) | Virus | S | Swine vesicular disease |
| Taenia spp. | Helminth | DCR | Taeniasis |
| Taylorella equigenitalis | Bacteria | E | Contagious Equine Metritis (CEM) |
| Togaviridae alphavirus | Virus | E | Encephalitis |
| Toxocara spp. | Nematode | DCR | Toxocariasis |
| Toxoplasma gondii | Protozoa | DCRS | Toxoplasma infection |
| Transmissible gastroenteritis virus | Virus | S | diarrhea |
| Tritrichomonas foetus | Protozoa | O | Trichomoniasis |
| Trichomonas gallinae | Protozoa | B | Trichomoniasis, cankers |
| Trichuriasis | Nematode | D | Whipworm infestation |
| Trixacarus caviae | Insect | R | infestation |
| Tritrichomonas foetus | Protozoa | DC | infestation |
| Trypanosoma cruzi | Protozoa | DCR | Trypanosomiasis |
| Venezuelan Equine Encephalomyelitis (VEE) | Virus | DRE | Encephalitis |
| Vesicular Stomatitis (VSV) | Virus | DREO | ulcers, lesions, fever |
| Wesselsbron Disease | Virus | DREO | flu-like illness |
| West Nile Virus | Virus | DCBRE | flu-like illness |
| Western Equine Encephalomyelitis (WEE) | Virus | DRE | Encephalitis |
| Brachyspira hyodysenteriae | Bacteria | S | dysentery |
| Campylobacter fetus | Bacteria | O | Campylobacteriosis |
| Corynebacterium bovis | Bacteria | R | hyperkeratosis |
| Corynebacterium kutscheri | Bacteria | R | pseudotuberculosis |
| Haemophilus parasuis | Bacteria | S | Glasser's Disease |
| Herpes virus BHV-1 (IBR virus) | Virus | O | Infectious bovine rhinotracheitis |
| Ibaraki virus | Virus | O | Ibaraki Disease |
| Junin virus | Virus | R | hemorrhagic fever |
| Klebsiella orthinolytica | Bacteria | R | pneumonia |
| Klebsiella oxytoca | Bacteria | R | pneumonia |
| Klebsiella planticola | Bacteria | R | pneumonia |
| Morbillivirus | Virus | O | Rinderpest |
| Mousepox | Virus | R | pox |
| Mycoplasma mycoides | Bacteria | O | Contagious bovine pleuropneumonia |
| Pestivirus | Virus | SO | Classical Swine Fever, BVD |
| Pneumonia Virus of Mice (PVM) | Virus | R | pneumonia |
| Pseudocowpox | Virus | O | viral skin disease |
| Rhadinovirus | Virus | O | Malignant catarrhal fever (MCF) |
| Sendai | Virus | R | Sendai disease |
| Staphylococcus hyicus | Bacteria | S | Epidermitis |
| Streptococcus canis | Bacteria | O | Streptococcosis |
| Streptococcus iniae | Bacteria | O | Streptococcosis |
| Streptococcus suis | Bacteria | SO | URD, Streptococcosis |

NOTES for Appendix A:

S = Swine

O = Cow, Bovine

D = Dog

C = Cat

P = Poultry (non-pet)

R = Rodent

E = Horses

B = Bird (pets)